

# (12) United States Patent

#### Andersen

#### US 9,273,297 B2 (10) **Patent No.:**

#### (45) **Date of Patent:** \*Mar. 1, 2016

#### (54) STABILIZATION OF ALPHA-AMYLASES TOWARDS CALCIUM DEPLETION AND ACIDIC PH

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(\*) Notice: Subject to any disclaimer, the term of this

patent is extended or adjusted under 35

U.S.C. 154(b) by 0 days.

This patent is subject to a terminal dis-

claimer.

(21) Appl. No.: 14/525,877

(22) Filed: Oct. 28, 2014

(65)**Prior Publication Data** 

> US 2015/0044732 A1 Feb. 12, 2015

#### Related U.S. Application Data

- (62) Division of application No. 13/518,962, filed as application No. PCT/EP2011/050074 on Jan. 4, 2011, now Pat. No. 8,900,848.
- (60) Provisional application No. 61/362,536, filed on Jul. 8, 2010, provisional application No. 61/355,230, filed on Jun. 16, 2010, provisional application No. 61/354,817, filed on Jun. 15, 2010, provisional application No. 61/354,775, filed on Jun. 15, 2010, provisional application No. 61/333,930, filed on May 12, 2010, provisional application No. 61/304,092, filed on Feb. 12, 2010, provisional application No. 61/292,327, filed on Jan. 5, 2010, provisional application No. 61/292,324, filed on Jan. 5, 2010.

#### (30)Foreign Application Priority Data

Jan. 4, 2010	(EP)	10150062
Jan. 4, 2010	(EP)	10150063

(51)	Int. Cl.	
	C12N 9/12	(2006.01)
	C07K 1/00	(2006.01)
	C12N 9/28	(2006.01)
	C11D 3/386	(2006.01)
	C12P 7/06	(2006.01)
	C12P 19/04	(2006.01)
	C12P 19/14	(2006.01)
	C12N 9/26	(2006.01)
	C12P 7/14	(2006.01)

(52) U.S. Cl.

CPC ...... C12N 9/2417 (2013.01); C11D 3/386 (2013.01); C12N 9/2414 (2013.01); C12P 7/06 (2013.01); C12P 7/14 (2013.01); C12P 19/04

(2013.01); C12P 19/14 (2013.01); C12Y **302/01001** (2013.01); Y02E 50/17 (2013.01)

(58) Field of Classification Search

CPC ...... C12N 9/2417; C12Y 302/01001; C12P 2201/00

See application file for complete search history.

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#### (57) **ABSTRACT**

The present invention relates to variants of a parent alphaamylase, the variant having improved stability or activity at low calcium conditions or at low pH.

#### 16 Claims, No Drawings

### STABILIZATION OF ALPHA-AMYLASES TOWARDS CALCIUM DEPLETION AND ACIDIC PH

#### CROSS-REFERENCE TO RELATED APPLICATIONS

This application is a division of U.S. application Ser. No. 13/518,962 filed Jun. 25, 2012 (now allowed), which is a 35 U.S.C. 371 national application of PCT/EP2011/050074 filed 10 Jan. 4, 2011 which claims priority or the benefit under 35 U.S.C. 119 of European application nos. 10150063.5 and 10150062.7 filed Jan. 4, 2010 and Jan. 4, 2010 and U.S. provisional application Nos. 61/292,324, 61/292,327, 61/304,092, 61/333,930, 61/354,775, 61/354,817, 61/355, 230 and 61/362,536 filed Jan. 5, 2010, Jan. 5, 2010, Feb. 12, 2010, May 12, 2010, Jun. 15, 2010, Jun. 15, 2010 Jun. 16, 2010 and Jul. 8, 2010 the contents of which are fully incorporated herein by reference.

#### REFERENCE TO SEQUENCE LISTING

This application contains a Sequence Listing in computer readable form, which is incorporated herein by reference.

#### FIELD OF THE INVENTION

The present invention relates to variants of an alpha-amylase having improved stability at an acidic pH and/or in the presence of strong chelators compared to its parent enzyme. 30 Further, the invention relates to nucleic acids encoding the variants, methods of producing the variants, and methods for using the variants.

#### BACKGROUND OF THE INVENTION

Alpha-amylases (alpha-1,4-glucan-4-glucanohydrolases, E.C. 3.2.1.1) constitute a group of enzymes, which catalyzes hydrolysis of starch and other linear and branched 1,4-gluosidic oligo- and polysaccharides.

There is a long history of industrial use of alpha-amylases in several known applications such as detergent, baking, brewing, starch liquefaction and saccharification, e.g., in preparation of high fructose syrups or as part of ethanol production from starch. These and other applications of 45 alpha-amylases are known and utilize alpha-amylases derived from microorganisms, in particular bacterial alpha-

Among the first bacterial alpha-amylases to be used was an alpha-amylase from B. licheniformis, also known as Ter- 50 mamyl, which has been extensively characterized and the crystal structure has been determined for this enzyme. Alkaline amylases, such as AA560 (SEQ ID NO: 2), disclosed in WO 00/60060, form a particular group of alpha-amylases that have found use in detergents. Many of these known bacterial 55 amylases have been modified in order to improve their functionality in a particular application.

Termamyl and many highly efficient alpha-amylases required calcium for activity. In the crystal structure for Teralpha-amylase structure coordinated by negatively charged amino acid residues. This requirement for calcium is a disadvantage in applications where strong chelating compounds are present, such as in detergents or during ethanol production from whole grains, where plant material comprising high 65 amount of natural chelaters such as phytate is hydrolysed using alpha-amylases.

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Calcium-insensitive amylases are known, e.g., the alphaamylases disclosed in EP 1022334 and WO 03/083054, and a Bacillus circulans alpha-amylase having the sequence disclosed in UNIPROT:Q03657, but these amylases are inferior to many of the calcium-sensitive amylase when it comes to starch hydrolysis and starch removal in various applications.

It would therefore be beneficial to provide variants of a calcium-sensitive alpha-amylase with reduced calcium sensitivity compared to its parent enzyme.

#### SUMMARY OF THE INVENTION

The present invention relates to isolated variants of a parent Termamyl-like alpha-amylase, comprising an alteration at two, three, four or five positions corresponding to positions 163, 188, 205, 208 and 209 of amino acids 1 to 485 of SEQ ID NO: 2 wherein the alteration(s) are independently

- (i) an insertion of an amino acid immediately downstream of the position,
- (ii) a deletion of the amino acid which occupies the position, and/or
- (iii) a substitution of the amino acid which occupies the position, and

wherein the variants have alpha-amylase activity. The variants of the invention may further comprise one or

more additional substitution(s).

Additionally, the isolated variants may comprise further alterations known to improve the performance of alpha-amylases including a deletion corresponding to amino acids 183 and 184 and substitutions in one or more of the positions 186, 193, 195, 202, 206, 214, 244, 452, 474 and 475, and each position corresponds to a position of the amino acid sequence of the enzyme having the amino acid sequence of SEQ ID NO: 2.

35 The variants of the invention have reduced calcium sensitivity compared with the parent alpha-amylase.

The present invention also relates to isolated nucleotide sequences encoding the variant alpha-amylases or polypeptides having alpha-amylase activity and to nucleic acid constructs, vectors, and host cells comprising the nucleotide sequences.

Methods for preparing the variants of the invention are also provided.

The present invention also relates to compositions comprising the variants of the invention, in particular a detergent additive composition, detergent composition, composition for manual or automatic dishwashing or compositions for manual or automatic laundry washing. Further, the invention relates to the use of an alpha-amylase variant according to the invention for washing and/or dishwashing, textile desizing and starch liquefaction. The invention also relates to a method for producing ethanol or other chemicals using the variant of the invention.

#### DETAILED DESCRIPTION OF THE INVENTION

#### Definitions

Alpha-Amylases (alpha-1,4-glucan-4-glucanohydrolases, mamyl it was found that four calcium atom were bound in the 60 E.C. 3.2.1.1) constitute a group of enzymes, which catalyze hydrolysis of starch and other linear and branched 1,4-glucosidic oligo- and polysaccharides.

> cDNA: The term "cDNA" means a DNA molecule that can be prepared by reverse transcription from a mature, spliced, mRNA molecule obtained from a eukaryotic cell. cDNA lacks intron sequences that may be present in the corresponding genomic DNA. The initial, primary RNA transcript is a

precursor to mRNA that is processed through a series of steps, including splicing, before appearing as mature spliced mRNA.

Coding sequence: The term "coding sequence" means a polynucleotide, which directly specifies the amino acid 5 sequence of its polypeptide product. The boundaries of the coding sequence are generally determined by an open reading frame, which usually begins with the ATG start codon or alternative start codons such as GTG and TTG and ends with a stop codon such as TAA, TAG, and TGA. The coding 10 sequence may be a DNA, cDNA, synthetic, or recombinant polynucleotide.

Control sequences: The term "control sequences" means all components necessary for the expression of a polynucle-otide encoding a variant of the present invention. Each control sequence may be native or foreign to the polynucleotide encoding the variant or native or foreign to each other. Such control sequences include, but are not limited to, a leader, polyadenylation sequence, propeptide sequence, promoter, signal peptide sequence, and transcription terminator. At a 20 minimum, the control sequences include a promoter, and transcriptional and translational stop signals. The control sequences may be provided with linkers for the purpose of introducing specific restriction sites facilitating ligation of the control sequences with the coding region of the polynucle-otide encoding a variant.

Expression: The term "expression" includes any step involved in the production of the variant including, but not limited to, transcription, post-transcriptional modification, translation, post-translational modification, and secretion.

Expression vector: The term "expression vector" means a linear or circular DNA molecule that comprises a polynucleotide encoding a variant and is operably linked to additional nucleotides that provide for its expression.

Fragment: The term "fragment" means a polypeptide hav- 35 ing one or more (several) amino acids deleted from the amino and/or carboxyl terminus of a mature polypeptide; wherein the fragment has alpha-amylase activity.

Host cell: The term "host cell" means any cell type that is susceptible to transformation, transfection, transduction, and 40 the like with a nucleic acid construct or expression vector comprising a polynucleotide of the present invention. The term "host cell" encompasses any progeny of a parent cell that is not identical to the parent cell due to mutations that occur during replication.

Improved pH stability: The term "improved pH stability" is defined herein as a variant enzyme displaying retention of enzymatic activity after a period of incubation at a particular pH, which reduces the enzymatic activity of the parent enzyme. Improved pH stability may also result in variants 50 better able to catalyze a reaction under such pH conditions.

Isolated variant: The terms "isolated" and "purified" mean a polypeptide or polynucleotide that is removed from at least one component with which it is naturally associated. For example, a variant may be at least 1% pure, e.g., at least 5% 55 pure, at least 10% pure, at least 20% pure, at least 40% pure, at least 60% pure, at least 80% pure, and at least 90% pure, as determined by SDS-PAGE and a polynucleotide may be at least 1% pure, e.g., at least 5% pure, at least 10% pure, at least 20% pure, at least 40% pure, at least 80% 60 pure, at least 90% pure, and at least 95% pure, as determined by agarose electrophoresis.

Mature polypeptide: The term "mature polypeptide" means a polypeptide in its final form following translation and any post-translational modifications, such as N-terminal 65 processing, C-terminal truncation, glycosylation, phosphorylation, etc. It is known in the art that a host cell may produce

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a mixture of two of more different mature polypeptides (i.e., with a different C-terminal and/or N-terminal amino acid) expressed by the same polynucleotide.

Nucleic acid construct: The term "nucleic acid construct" means a nucleic acid molecule, either single- or double-stranded, which is isolated from a naturally occurring gene or is modified to contain segments of nucleic acids in a manner that would not otherwise exist in nature or which is synthetic. The term nucleic acid construct is synonymous with the term "expression cassette" when the nucleic acid construct contains the control sequences required for expression of a coding sequence of the present invention.

Operably linked: The term "operably linked" means a configuration in which a control sequence is placed at an appropriate position relative to the coding sequence of a polynucle-otide such that the control sequence directs the expression of the coding sequence.

Parent Enzyme: The term "parent" alpha-amylase as used herein means an alpha-amylase to which modifications, e.g., substitution(s), insertion(s), deletion(s), and/or truncation(s), are made to produce the enzyme variants of the present invention. This term also refers to the polypeptide with which a variant is compared and aligned. The parent may be a naturally occurring (wild type) polypeptide, or it may be a variant thereof, prepared by any suitable means. For instance, the parent protein may be a variant of a naturally occurring polypeptide which has been modified or altered in the amino acid sequence. A parent may also be an allelic variant which is a polypeptide encoded by any of two or more alternative forms of a gene occupying the same chromosomal locus.

Sequence Identity: The relatedness between two amino acid sequences or between two nucleotide sequences is described by the parameter "sequence identity".

For purposes of the present invention, the degree of sequence identity between two amino acid sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443-453) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, *Trends Genet.* 16: 276-277), preferably version 3.0.0 or later. The optional parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EBLO-SUM62 (EMBOSS version of BLOSUM62) substitution matrix. The output of Needle labeled "longest identity" (obtained using the—nobrief option) is used as the percent identity and is calculated as follows:

(Identical Residues×100)/(Length of Alignment-Total Number of Gaps in Alignment)

For purposes of the present invention, the degree of sequence identity between two deoxyribonucleotide sequences is determined using the Needleman-Wunsch algorithm (Needleman and Wunsch, 1970, supra) as implemented in the Needle program of the EMBOSS package (EMBOSS: The European Molecular Biology Open Software Suite, Rice et al., 2000, supra), preferably version 3.0.0 or later. The optional parameters used are gap open penalty of 10, gap extension penalty of 0.5, and the EDNAFULL (EMBOSS version of NCBI NUC4.4) substitution matrix. The output of Needle labeled "longest identity" (obtained using the—nobrief option) is used as the percent identity and is calculated as follows:

(Identical Deoxyribonucleotidesx100)/(Length of Alignment-Total Number of Gaps in Alignment)

Subsequence: The term "subsequence" means a polynucleotide having one or more (several) nucleotides deleted from

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the 5'- and/or 3'-end of a mature polypeptide coding sequence; wherein the subsequence encodes a fragment having alpha-amylase activity.

Variant: The term "variant" is defined herein as an alphaamylase comprising one or more alterations, such as substitutions, insertions, deletions, and/or truncations of one or more specific amino acid residues at one or more specific positions in the polypeptide.

Wild-Type Enzyme: The term "wild-type" alpha-amylase denotes an alpha-amylase expressed by a naturally occurring microorganism, such as an yeast or filamentous fungus found

#### Conventions for Designation of Variants

In the present invention, a specific numbering of amino 15 acid residue positions in the alpha-amylase variants is employed. For example, by aligning the amino acid sequences of known alpha-amylases, it is possible to designate an amino acid position number to any amino acid residue in any alpha-amylase enzyme.

Using the numbering system originating from the amino acid sequence of the alpha-amylase disclosed in SEQ ID NO: 2, aligned with the amino acid sequence of a number of other alpha-amylases, it is possible to indicate the position of an amino acid residue in an alpha-amylase in regions of struc- 25 tural homology.

In describing the various alpha-amylase variants of the present invention, the nomenclature described below is adapted for ease of reference. In all cases, the accepted employed.

#### Substitutions.

For an amino acid substitution, the following nomenclature is used: Original amino acid, position, substituted amino acid. Accordingly, the substitution of threonine with alanine at 35 position 226 is designated as "Thr226Ala" or "T226A". Multiple mutations are separated by addition marks ("+"), e.g., "Gly205Arg+Ser411Phe" or "G205R+S411F", representing mutations at positions 205 and 411 substituting glycine (G) with arginine (R), and serine (S) with phenylalanine (F), 40 respectively. In case that an amino acid substitution in a particular position is specified where the position can be occupied by different amino acids depending of the actual parent the original amino acid is indicated as X or Xaa. For example "X226A" is intended to mean that the amino acid 45 that occupies position 226 is substituted with A or Ala, independently of which amino acid occupies position 226 in the original sequence (parent sequence).

#### Deletions.

For an amino acid deletion, the following nomenclature is 50 used: Original amino acid, position\*. Accordingly, the deletion of glycine at position 195 is designated as "Gly195\*" or "G195\*". Multiple deletions are separated by addition marks ("+"), e.g., "Gly195\*+Ser411\*" or "G195\*+S411\*". Insertions.

For an amino acid insertion, the following nomenclature is used: Original amino acid, position, original amino acid, new inserted amino acid. Accordingly the insertion of lysine after glycine at position 195 is designated "Gly195GlyLys" or "G195GK". Multiple insertions of amino acids are desig- 60 nated [Original amino acid, position, original amino acid, new inserted amino acid #1, new inserted amino acid #2; etc.]. For example, the insertion of lysine and alanine after glycine at position 195 is indicated as "Gly195GlyLysAla" or "G195GKA".

In such cases the inserted amino acid residue(s) are numbered by the addition of lower case letters to the position 6

number of the amino acid residue preceding the inserted amino acid residue(s). In the above example the sequences would thus be:

,	Parent:	Variant:
	195	195 195a 195b
0	G	G - K - A

#### Degenerate Indications.

For degenerate indications where an amino acid residue identical to the existing amino acid residue is inserted, degeneracy in the nomenclature arises. For example, a glycine inserted after the glycine in the above example would be indicated by "G195GG". Given that an alanine were present at position 194, the same actual change could just as well be indicated as "A194AG":

		Parent:	Variant:	
	Numbering I:	194 195	194 195 195a	
5	Sequence:	A - G	A - G - G	
	Numbering II:		194 194a 195	

Such instances will be apparent to the skilled person, and IUPAC single letter or triple letter amino acid abbreviation is 30 the indication "G195GG" and corresponding indications for this type of insertion is thus meant to comprise such equivalent degenerate indications.

> If amino acid sequence segments are repeated in the parent polypeptide and/or in the variant, equivalent degenerate indications arise, also when alterations other than insertions are listed such as deletions and/or substitutions. For example, the deletion of two consecutive amino acids "AG" in the sequence "AGAG" from position 194-97 may be written as "A194\*+G195\*" or "A196\*+G197\*":

	Parent:	Variant:
Numbering I:	194 195 196 197	194 195
Sequence:	A - G - A - G	A - G
Numbering II:		196 197

#### Multiple Modifications.

Variants comprising multiple modifications are separated by addition marks ("+"), e.g., "Arg170Tyr+Gly195Glu" or "R170Y+G195E" representing modifications at positions 170 and 195 substituting tyrosine and glutamic acid for arginine and glycine, respectively. Thus, "Tyr167Gly,Ala,Ser, Thr+Arg170Gly,Ala,Ser,Thr" designates the following vari-

"Tyr167Gly+Arg170Gly", "Tyr167Gly+Arg170Ser",

'Tyr167Gly+Arg170Thr", "Tyr167Ala+Arg170Ala", "Tyr167Ala+Arg170Ser",

"Tyr167Ser+Arg170Gly" "Tyr167Ser+Arg170Ala",

"Tyr167Ser+Arg170Thr",

"Tyr167Thr+Arg170Gly", "Tyr167Thr+Arg170Ser", and "Tyr167Thr+Arg170Thr".

"Tyr167Gly+Arg170Ala",

"Tyr167Ala+Arg170Gly",

"Tyr167Ala+Arg170Thr",

"Tyr167Ser+Arg170Ser",

"Tyr167Thr+Arg170Ala",

This nomenclature is particularly relevant to modifications involving substituting, inserting or deleting amino acid residues having specific common properties. Such modifications are referred to as conservative amino acid modification(s). Examples of conservative modifications are within the group of basic amino acids (arginine, lysine and histidine), acidic amino acids (glutamic acid and aspartic acid), polar amino acids (glutamine and asparagine), hydrophobic amino acids (leucine, isoleucine and valine), aromatic amino acids (phenylalanine, tryptophan and tyrosine), and small amino acids (glycine, alanine, serine, threonine and methionine). Amino acid modifications, which do not generally alter the specific activity are known in the art and are described, for example, by H. Neurath and R. L. Hill, 1979, In, The Proteins, Academic Press, New York. The most commonly occurring exchanges are Ala/Ser, Val/Ile, Asp/Glu, Thr/Ser, Ala/Gly, Ala/Thr, Ser/Asn, Ala/Val, Ser/Gly, Tyr/Phe, Ala/Pro, Lys/ Arg, Asp/Asn, Leu/Ile, Leu/Val, Ala/Glu, and Asp/Gly as well as the reverse (Taylor, 1986, Journal of Theoretical Biology 205-218: 119: compbio.dundee.ac.uk/papers/amas/ 20 amas3d.html).

#### Parent Alpha-Amylases

The parent alpha-amylase may in principle be any alphaamylase for which it is desired to prepare a variant having improved stability at low pH. Alpha-amylases are known 25 derived from a vide selection of organism including bacteria, such as from species of the genus Bacillus, e.g., Bacillus licheniformis; from species of fungi, such as Aspergillus oryzae (TAKA-amylase) or Aspergillus niger; from plants such as barley and from mammals. The parent alpha-amylase 30 may in principle be any such alpha-amylase irrespective of the origin.

## Termamyl-Like Alpha-Amylases

It is well known that a number of alpha-amylases produced by Bacillus spp. are highly homologous on the amino acid 35 level as well as on the structural level. For instance, the B. licheniformis alpha-amylase comprising the amino acid sequence shown in SEQ ID NO: 4 (commercially available as Termamyl<sup>TM</sup>) has been found to be about 81% homologous amino acid sequence shown in SEQ ID NO: 6 and about 60% homologous with the B. stearothermophilus alpha-amylase comprising the amino acid sequence shown in SEQ ID NO: 8. Further homologous alpha-amylases include an alpha-amylase derived from a strain of the Bacillus sp. NCIB 12289, 45 NCIB 12512, NCIB 12513 or DSM 9375, all of which are described in detail in WO 95/26397, and the SP#707 alphaamylase described by Tsukamoto et al., 1988, Biochemical and Biophysical Research Communications 151: 25-31.

Still further homologous alpha-amylases include the 50 alpha-amylase produced by the B. licheniformis strain described in EP 0252666 (ATCC 27811), and the alphaamylases identified in WO 91/00353 and WO 94/18314. Other commercial Termamyl-like alpha-amylases are comprised in the products sold under the following tradenames: 55 Optitherm<sup>TM</sup> and Takatherm<sup>TM</sup> (available from Solvay); Maxamyl<sup>TM</sup> (available from Gist-brocades/Genencor), Spezym AA<sup>TM</sup> and Spezyme Delta AAT<sup>TM</sup>, Spezyme FRED (available from Genencor), and Keistase™ (available from Daiwa), Purastar<sup>TM</sup> ST 5000E, PURASTRA<sup>TM</sup> HPAM L 60 (from Genencor Int.).

Because of the substantial homology found between these alpha-amylases, they are considered to belong to the same class of alpha-amylases, namely the class of "Termamyl-like alpha-amylases".

Accordingly, in the present context, the term "Termamyllike alpha-amylase" is intended to indicate an alpha-amylase,

which at the amino acid level exhibits a substantial homology to Termamyl<sup>TM</sup>, i.e., the B. licheniformis alpha-amylase having the amino acid sequence shown in SEQ ID NO: 4. In other words, a Termamyl-like alpha-amylase is an alpha-amylase, which has the amino acid sequence shown in SEQ ID NO: 2, 4, or 6, and the amino acid sequence shown in SEQ ID NO: 1 or 2 of WO 95/26397 or in Tsukamoto et al. (1988), or the Bacillus flavothermus amylase, AMY1048 described in WO 2005/001064, or the alpha-amylase TS-22 having the amino acid sequence of SEQ ID NO: 12; or the alpha-amylase TS-23 having the amino acid sequence of SEQ ID NO: 13, described in J. Appl. Microbiology, 1997, 82: 325-334 (SWALL: q59222), or the alpha-amylase derived from Bacillus sp. KSM-AP1378 (FERM BP-3048) having the amino acid sequence of SEQ ID NO: 14, described in WO 97/00324, or the alpha-amylase derived from Bacillus sp. A 7-7 having the amino acid sequence of SEQ ID NO: 15, described in WO 02/10356 or the Cytophaga alpha-amylase having the amino acid sequence of SEQ ID NO: 16, described in Jeang et al., 2002, Appl. Environ. Microbiol. 68:3651-3654, or the alphaamylase derived from Bacillus stearothermophilus (Spezyme Xtra), having the amino acid sequence of SEQ ID NO: 17; or the alpha-amylase produced by the B. licheniformis strain described in EP 0252666 (ATCC 27811) or the alpha-amylases disclosed in WO 91/00353 and WO 94/18314 or i) which displays at least 60%, preferred at least 70%, more preferred at least 75%, even more preferred at least 80%, especially at least 85%, especially preferred at least 90%, even especially more preferred at least 95% homology, more preferred at least 97%, more preferred at least 99% with at least one of said amino acid sequences and/or ii) displays immunological cross-reactivity with an antibody raised against at least one of said alpha-amylases, and/or iii) is encoded by a DNA sequence which hybridises to the DNA sequences encoding the above-specified alpha-amylases which are apparent from SEQ ID NOS: 1, 3, and 5 of the present application and SEQ ID NOS: 4 and 5 of WO 95/26397, respectively.

In a preferred embodiment the parent Termamyl-like alpha with the B. amyloliquefaciens alpha-amylase comprising the 40 amylase is SEQ ID NO: 10 (SEQ ID NO: 2 of WO 95/26397), SEQ ID NO: 8 or SEQ ID NO: 2 including any of [SEQ ID NO: 10]+R181\*+G182\*, [SEQ ID NO: 10]+D183\*+G184\*; [SEQ ID NO: 10]+D183\*+G184\*+N195F; [SEQ ID NO: 10]+D183\*+G184\*+M202L; [SEQ ID NO: 10]+D183\*+ G184\*+N195F+M202L; [SEQ ID NO: 10]+D183\*+G184\*+ R181Q; [SEQ ID NO: 10]+D183\*+G184\*+R118K+N195F+ R320K+R458K; [SEQ ID NO: 8]+I181\*+G182\*; [SEQ ID NO: 8]+I181\*+G182\*+N193F; [SEQ ID NO: 8]+I181\*+ G182\*+M200L; [SEQ ID NO: 8]+I181\*+G182\*+N193F+ M200L; [SEQ ID NO: 10]+D183\*+G184\*; [SEQ ID NO: 10]+D183\*+G184\*+N195F; [SEQ ID NO: 10]+D183\*+ G184\*+M202L; [SEQ ID NO: 10]+D183\*+G184\*+N195F+ M202L; [SEQ ID NO: 10]+D183\*+G184\*+R118K+ N195F+R320K+R458K.

[SEQ ID NO: 8]+I181\*+G182\*+N193F" means the B. stearothermophilus alpha-amylase having SEQ ID NO: 8 has been mutated as follows: deletions in positions I181 and G182 and a substitution from Asn (N) to Phe (F) in position 193.

#### Parent Hybrid Alpha-Amylases

The parent alpha-amylase may be a hybrid alpha-amylase, i.e., an alpha-amylase, which comprises a combination of partial amino acid sequences derived from at least two alphaamylases.

The parent hybrid alpha-amylase may be one, which on the basis of amino acid homology and/or immunological crossreactivity and/or DNA hybridization (as defined above) can

be determined to belong to the Termamyl-like alpha-amylase family. In this case, the hybrid alpha-amylase is typically composed of at least one part of a Termamyl-like alpha-amylase and part(s) of one or more other alpha-amylases selected from Termamyl-like alpha-amylases or non-Termamyl-like alpha-amylases of microbial (bacterial or fungal) and/or mammalian origin.

Thus, the parent hybrid alpha-amylase may comprise a combination of partial amino acid sequences deriving from at least two Termamyl-like alpha-amylases, or from at least one 10 Termamyl-like and at least one non-Termamyl-like bacterial alpha-amylase, or from at least one Termamyl-like and at least one fungal alpha-amylase. The Termamyl-like alpha-amylase, from which a partial amino acid sequence derives, may, e.g., be any of those specific Termamyl-like alpha-amylases referred to herein.

For instance, the parent alpha-amylase may comprise a C-terminal part of an alpha-amylase derived from a strain of B. licheniformis, and a N-terminal part of an alpha-amylase derived from a strain of B. amyloliquefaciens or from a strain 20 of B. stearothermophilus. For instance, the parent alpha-amylase may comprise at least 430 amino acid residues of the C-terminal part of the B. licheniformis alpha-amylase, and may, e.g., comprise a) an amino acid segment corresponding to the 37 N-terminal amino acid residues of the B. amv- 25 loliquefaciens alpha-amylase having the amino acid sequence shown in SEQ ID NO: 6 and an amino acid segment corresponding to the 445 C-terminal amino acid residues of the B. licheniformis alpha-amylase having the amino acid sequence shown in SEQ ID NO: 4, or b) an amino acid 30 segment corresponding to the 68 N-terminal amino acid residues of the B. stearothermophilus alpha-amylase having the amino acid sequence shown in SEQ ID NO: 8 and an amino acid segment corresponding to the 415 C-terminal amino acid residues of the B. licheniformis alpha-amylase having the 35 amino acid sequence shown in SEQ ID NO: 4.

In a preferred embodiment the parent Termamyl-like alpha-amylase is a hybrid Termamyl-like alpha-amylase identical to the *Bacillus licheniformis* alpha-amylase shown in SEQ ID NO: 4, except that the N-terminal 35 amino acid 40 residues (of the mature protein) is replaced with the N-terminal 33 amino acid residues of the mature protein of the *Bacillus amyloliquefaciens* alpha-amylase (BAN) shown in SEQ ID NO: 6. Said hybrid may further have the following mutations: H156Y+A181T+N190F+A209V+Q264S (using the 45 numbering in SEQ ID NO: 4) referred to as LE174.

The non-Termanyl-like alpha-amylase may, e.g., be a fungal alpha-amylase, a mammalian or a plant alpha-amylase or a bacterial alpha-amylase (different from a Termanyl-like alpha-amylase). Specific examples of such alpha-amylases 50 include the *Aspergillus oryzae* TAKA alpha-amylase, the *A. niger* acid alpha-amylase, the *Bacillus subtilis* alpha-amylase, the porcine pancreatic alpha-amylase and a barley alpha-amylase. All of these alpha-amylases have elucidated structures, which are markedly different from the structure of a 55 typical Termanyl-like alpha-amylase as referred to herein.

The fungal alpha-amylases mentioned above, i.e., derived from *A. niger* and *A. oryzae*, are highly homologous on the amino acid level and generally considered to belong to the same family of alpha-amylases. The fungal alpha-amylase 60 derived from *Aspergillus oryzae* is commercially available under the tradename Fungamyl<sup>TM</sup>.

Furthermore, when a particular variant of a Termamyl-like alpha-amylase (variant of the invention) is referred to—in a conventional manner—by reference to modification (e.g., 65 deletion or substitution) of specific amino acid residues in the amino acid sequence of a specific Termamyl-like alpha-amy-

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lase, it is to be understood that variants of another Termamyllike alpha-amylase modified in the equivalent position(s) (as determined from the best possible amino acid sequence alignment between the respective amino acid sequences) are encompassed thereby.

A preferred variant of the invention is one derived from a *B. licheniformis* alpha-amylase (as parent Termamyl-like alpha-amylase), e.g., one of those referred to above, such as the *B. licheniformis* alpha-amylase having the amino acid sequence shown in SEQ ID NO: 4.

Variants

The present invention relates to isolated variant alphaamylases of a parent Termamyl-like alpha-amylase comprising two, three, four or five amino acid alterations in positions corresponding to positions in the parent alpha-amylase selected from the group consisting of 163, 188, 205, 208 and 209; the alteration(s) are independently

- (i) an insertion of an amino acid immediately downstream of the position,
- (ii) a deletion of the amino acid which occupies the position, and/or
- (iii) a substitution of the amino acid which occupies the position,

wherein the variant has alpha-amylase activity and have reduced calcium sensitivity compared with the parent alpha-amylase, and wherein each position corresponds to a position of the amino acid sequence of the enzyme having the amino acid sequence of SEQ ID NO: 2.

The variant may be a natural or a non-natural variant, where natural variants should be understood as an alphaamylase isolated from a naturally occurring organism that have not been the subject of human manipulation. A non-natural variant is a variant that have been modified from its natural counterpart, the parent alpha-amylase, by human intervension using techniques such as mutation of a wild type organism and isolation of variant alpha-amylases, techniques involving isolation and manipulation of nucleic acids encoding a parent alpha-amylase or chemical synthesis of the variants of nucleic acids encoding them. Many such techniques are available in the art and the skilled person will appreciate that such techniques can be applied in the present invention.

The variants of the invention are generally isolated using at least one separation step, and the invention does therefore not apply to natural enzymes in their natural environment.

The variants according to the invention have the benefit of being less sensitive toward calcium depletion than their parent alpha-amylase, but at the same time they have maintained the performance properties of the parent alpha-amylase. Calcium sensitivity is manifested in the activity and/or stability of the particular alpha-amylase in calcium-depleted environments and/or under acidic conditions. Calcium-depleted environments occur in many known applications for alphaamylases, such as in the presence of strong chelators binding metal ions, in particular calcium ions, e.g., in detergents, where it is common to include strong chelators because of the beneficial effect of the laundering process, or in conditions where plant material including natural chelators such as phytates or citrates is present. Such strong chelators will compete for the calcium ions and will to some extend be able to deprive calcium-sensitive alpha-amylases for the calcium ions bound in their structure with the consequence that the stability or activity of the calcium sensitive alpha-amylase is reduced.

Acidic conditions may also affect the stability or activity of calcium-sensitive alpha-amylases. It is believed that low pH may lead to a protonation of the amino acid residues that coordinates the calcium ions in sensitive alpha-amylases with

the result that they no longer is capable of binding the calcium and the result is a loss of stability and/or activity. As examples of applications where alpha-amylases are exposed to acidic conditions can be mentioned use of alpha-amylases as in treatment of digestive disorders such as disclosed in WO 5 2006/136161, and use in feed.

Thus, the variants of the invention have at least one of the properties: improved stability and/or activity in the presence of strong chelators and/or improved stability and/or activity at low pH, and it should be understood in this specification and claims that a variant having reduced calcium sensitivity has improved stability and/or activity in the presence of strong chelators and/or improved stability and/or activity at low pH.

Chelator strength may be evaluated using methods known in the art such as methods disclosed in *Anal. Biochem.* 314: 15 227-234 (2003), and *JAOCS* 61(9): 1475-1478 (1984). As examples of strong chelators that may be used for such an assay can be mentioned EGTA (ethylene glycol tetraacetic acid), EDTA (ethylene diamine tetraacetic acid), DTPA (diethylene triamine pentaacetic acid), DTMPA (diethylene triamine-penta-methylene phosphonic acid) and HEDP (1-hydroxyethan-1,1-diylbis(phosphonic acid)). The skilled person will be able to select other strong chelators that may be used in determining the calcium sensitivity of an alpha-amylase

In the present invention, the isolated variants of a parent termamyl-like alpha-amylase comprise an alteration at two, three, four or five positions, said positions corresponding to positions in the parent alpha-amylase selected from the group consisting of: 163, 188, 205, 208 and 209 wherein the alteration(s) are independently

- (i) an insertion of an amino acid immediately downstream of the position,
- (ii) a deletion of the amino acid which occupies the position, and/or
- (iii) a substitution of the amino acid which occupies the position,

wherein the variant has alpha-amylase activity; and each position corresponds to a position of the amino acid sequence of the enzyme having the amino acid sequence of SEQ ID 40 NO: 2.

Preferably the variants comprises alterations at three positions, more preferred four positions even more preferred five positions and most preferred six positions, said positions corresponding to positions in the parent alpha-amylase 45 selected from the group consisting of: 163, 188, 205, 208 and 209 using SEQ ID NO: 2 for numbering.

The alterations may be selected among:

X163A,C,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y, preferably X163Q or X163N;

X188A,C,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y, preferably X188N;

X205A,C,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y, preferably X205N;

X208A,C,D,E,F,G,H,I,K,L,N,P,Q,R,S,T,V,W,Y, prefer- 55 ably X208F or X208Y; and

 $\dot{X}209A,C,E,F,G,H,I,K,L,M,N,P,Q,R,S,T,V,W,Y,$  preferably X209S or X209N.

Preferred variants comprise the mutations D163Q+ D188N+D209N,S or the mutations D163Q, N+D188N+ 60 M208F+D209S+K242P+S244W.

The variants may further comprise additional alteration(s) in one or more amino acid residues of the parent alphaamylase.

Preferred further alterations include alterations where one 65 or more amino acids in the B-domain of the parent alphaamylase are substituted with the corresponding amino acids

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of *B. circulans* alpha-amylase having SEQ ID NO: 9 or *Bacillus* sp. KSM-K38 alpha-amylase SEQ ID NO: 11. In case that a particular amino acid residue in the B-domain of a parent alpha-amylase is missing in SEQ ID NO: 9 or SEQ ID NO: 11 the preferred alteration is a deletion of the particular amino acid. The skilled person will appreciate that such a substitution of an amino acid residue in the B-domain of the parent alpha-amylase with the corresponding amino acid in SEQ ID NO: 9 or SEQ ID NO: 11 is only relevant for positions where the two amino acid sequences differ. Consequently, the number of possible alterations in this group will vary depending on the particular parent alpha-amylase and the sequence identity between the parent alpha-amylase and the alpha-amylase having the sequence shown in SEQ ID NO: 9 or SEQ ID NO: 11

The inventors have found that such further alterations provides for variants having an even more reduced calcium sensitivity compared with the parent alpha-amylase.

The B-domain of alpha-amylases is well known in the art and methods for identifying B-domains known in the art can also be applied to the present invention. The B-domain of alpha-amylases can be determined by structure analysis such as by using crystallographically techniques for identification of the domain structure of a given alpha-amylase. For example, Machius et al., 1995, J. Mol. Biol. 4: 545-559 and Machius et al., 1998, Structure 6:281-292, identified the B-domain in B. licheniformis alpha-amylase as residues 104-204; Brzozowski et al., 2000, Biochemistry 39:9099-9017 identified the B-domain in a hybrid alpha-amylase consisting of amino acids 1-300 of B. amyloliquefaciens alpha-amylase and amino acid residues 301-483 of B. licheniformis alphaamylase as residues 104-205; Suvd et al., 2001, J. Biochem. 129:461-468, identified the B-domain in B. stearopthermophilus alpha amylase and Nonaka et al. (2008) identified the 35 B-domain of the alpha-amylase from Bacillus sp. KSM-K38.

An alternative method for determining the B-domain for a given alpha-amylase is by sequence alignment of the amino acid sequence of the given alpha-amylase and an alpha-amylase for which the B-domain has been determined. The two sequences are aligned and the sequence in the given alphaamylase sequence that aligns with the B-domain sequence in the alpha-amylase for which the B-domain has been determined can for the purpose of this invention be considered the B-domain for the given alpha-amylase. This method is particular suitable for alpha-amylases for which the three-dimensional structure is not available. However, for alpha-amylases where the B-domain has been determined based on the three-dimensional structure of the alpha-amylase, the B-domain determined by the latter method should preferentially be used in case that the B-domain determined by alignment differs from the B-domain determined based on the structure.

The variants of the invention may even comprise further alterations known in the art to improve the performance of alpha-amylases. For example may oxidizable amino acid residues be substituted with a non-oxidizable amino acid residue in order to improve the stability of the enzyme under oxidizing conditions, e.g., in the presence of bleach, in accordance with the teachings of WO 94/18314 and WO 94/02597, incorporated herein by reference.

A two amino acid deletion may be introduced in positions corresponding to R181+G182 or T183+G184 in SEQ ID NO: 2 in accordance with the teachings of WO 96/23873, incorporated by reference.

Further beneficial substitutions that may be introduced into the variants of the invention can be found in WO 99/23211, WO 01/66712, WO 02/10355 and WO 2006/002643 all included by reference).

As examples of preferred further alterations can be mentioned D183\*+G184\*, G186A,Y,T, T193F, N195F, M202L, I,T,S,A, I206F,Y, V214I, S244A,D,E,N,Q,W, T452H,Y, G474R, G475R, wherein each position corresponds to a position of the amino acid sequence of the enzyme having the amino acid sequence of SEQ ID NO: 2. The skilled person will appreciate that corresponding alteration can be identified and performed starting from other parent alpha-amylases.

The number of amino acid substitutions in the variants of the present invention comprise preferably less than 60 substitutions, more preferred less than 55 substitutions, more preferred less than 45 substitutions, more preferred less than 45 substitutions, more preferred less than 40 substitutions, more preferred less than 35 substitutions, more preferred less than 25 substitutions, more preferred less than 25 substitutions, more preferred less than 25 substitutions, more preferred less than 15 substitutions, most preferred less than 10 substitutions.

The variants of the invention are preferably at least 70% 20 identical to their parent alpha-amylase, more preferred at least 75% identical to their parent alpha-amylase; more preferred at least 80% identical to their parent alpha-amylase, more preferred at least 85% identical to their parent alpha-amylase more preferred at least 90% identical to their parent 25 alpha-amylase more preferred at least 95% identical to their parent alpha-amylase, and most preferred at least 98% identical to their parent alpha-amylase.

In another embodiment, the variant has at least 70%, e.g., at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity to the mature polypeptide of SEQ ID NO: 2.

In another embodiment, the variant has at least 70%, e.g., at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity to the mature 40 polypeptide of SEO ID NO: 4.

In another embodiment, the variant has at least 70%, e.g., at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 45 99%, but less than 100%, sequence identity to the mature polypeptide of SEQ ID NO: 6.

In another embodiment, the variant has at least 70%, e.g., at least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, 50 such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity to the mature polypeptide of SEQ ID NO: 8.

In another embodiment, the variant has at least 70%, e.g., at least 75%, at least 80%, at least 85%, at least 90%, at least 55 91%, at least 92%, at least 93%, at least 94%, at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity to the mature polypeptide of SEQ ID NO: 10.

In another embodiment, the variant has at least 70%, e.g., at 60 least 75%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, such as at least 96%, at least 97%, at least 98%, and at least 99%, but less than 100%, sequence identity to the mature polypeptide of SEQ ID NO: 11.

In on preferred embodiment the parent alpha-amylase is the alpha-amylase having the sequence disclosed in SEQ ID 14

NO: 2. In this embodiment preferred substitutions according to the invention are selected among: D163Q,N, D188N, D205N, M208Y and D209N.

Alterations in the B-domain of SEQ ID NO: 2 include the alterations: A113E, M116V, V117F, R118K, A119V, V120I, N123D, N126D, N128T, Q129K, G133E, D134P, Y135F, T136E, A139G, D144T, N150D, T151Q, H152Y, N154S, R158N, W159S, Y160E, V165T, W167F, Q169A, S170R/K, R171E/G, K172E, L173R, N174\*, N175T, R176G, I177V, Y178F, K179R, F180I, R181A, D183E, G184N, A186K, W189E, E190N, T193D, N195F, Y203F, E212D, V214R, and N215R.

As examples of further alterations known to increase performance of alpha-amylases can be mentioned: D183\*+5 G184\*, G186A,Y or T, T193F, N195F, M202X preferably L,I,T,S or A, I206F or Y, V214I, S244A,D,E,N,Q or W, T452H or Y, G474R and G475R.

Preferred variants according to this embodiment include: D163Q+D188N+M208F+D209S+K242P+S244W;

- 0 D163N+R181A+G182A+K185T+G186N+D188N+ D205N+M208F+D209S+V238I+K242P+S244W; N128W+D163N+R181A+G182N+K185T+G186N+ D188N+D205N+M208F+D209S+K242P+S244W; D163N+R181A+G183N+K185T+G186N+D188N+
- D205N+M208F+D209S+K242P+S244W+H408W+ N409D+D432N+A434P; D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+N409D+ D432N+A434P;
- 30 D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W D183\*+G184\*+R118K+N195F+R320K+R458K+D163Q+ D188N+M208F+D209S+K242P+S244W; D183\*+G184\*+R118K+N195F+R320K+R458K+D163N+
- 5 R181A+G182A+K185T+G186N+D188N+D205N+ M208F+D209S+V238I+K242P+S244W; D183\*+G184\*+R118K+N195F+R320K+R458K+N128W+ D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W;
- 40 D183\*+G184\*+R118K+N195F+R320K+R458K+D163N+ R181A+G183N+K185T+G186N+D188N+D205N+ M208F+D209S+K242P+S244W+H408W+N409D+ D432N+A434P;
  - D183\*+G184\*+R118K+N195F+R320K+R458K;
- 45 D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W; D183\*+G184\*+R118K+N195F+R320K+R458K+D163N+ R181A+G182A+K185T+G186N+D188N+D205N+ M208F+D209S+V238I+K242P+S244W;
- D188N+D209S;
- D163N+D188N+D209S;
- D163N+D188N+D205N+D209S;
- D163N+D188N+D205N+M208F+D209S;
- D207N+D209S;
- 5 D163N+D207N+D209S;
- D163N+D188N+D207N+D209S;
- D163N+D188N+D199N+D207N+D209S;
- D163N+D188N+D199N+D205N+D207N+D209S;
- D163N+D188N+D199N+D205N+M208F+D207N+
- 0 D209S;
  - D163N+R181A+G182N+G186N+D188N+D205N+D209S; D163N+R181A+G182N+K185T+G186N+D188N+ D205N+D209S;
- D163N+R181A+G182N+K185T+G186N+D188N+
- D205N+M208F+D209S;
- D163N+R181A+G182N+D188N+D199N+D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+D188N+D199N+D205N+M208F+D207N+D209S; and D163N+R181A+G182N+K185T+G186N+D188N+D199N+D205N+M208F+D207N+D209S.

In another preferred embodiment the parent alpha-amylase is SP722 having the sequence disclosed in SEQ ID NO: 10. In this embodiment preferred substitutions according to the invention are selected among: D163Q,N, D188N, D205N, M208Y and D209N.

Alterations in the B-domain of SEQ ID NO: 10 include the alterations: A113E, N116V, V117F, L118K, A119V, V120I, N123D, N126D, N128T, Q129K, G133E, D134P, Y135F, T136E, A139G, D144T, N150D, T151Q, D154S, R158N, W159S, Y160E, V165T, W167F, Q169A, S170R or K, R171E or G, Q172E, F173R, Q174\*, N175T, R176G, I177V, Y178F, K179R, F180I, R181A, D183E, G184N, A186K, W189E, E190N, S193D, N195F, Y203F, V206I, E212D, V214R, and N215R.

As examples of further alterations known to increase performance of alpha-amylases can be mentioned: D183\*+G184\*, A186Y or T, S193F, N195F, M202X, preferably L,I, T,S or A, I206L or F, V214I, S244A,E or Q, H452Y, G474R and G475R.

As examples or preferred variants according to this <sup>25</sup> embodiment can be mentioned:

D188N+D209S;

D163N+D188N+D209S;

D163N+D188N+D205N+D209S:

D163N+D188N+D205N+M208F+D209S;

D207N+D209S;

D163N+D207N+D209S;

D163N+D188N+D207N+D209S;

D163N+D188N+D199N+D207N+D209S;

D163N+D188N+D199N+D205N+D207N+D209S;

D163N+D188N+D199N+D205N+M208F+D207N+D209S:

D163N+R181A+G182N+G186N+D188N+D205N+D209S; D163N+R181A+G182N+K185T+A186N+D188N+

D205N+D209S; D163N+R181A+G182N+K185T+A186N+D188N+

D205N+M208F+D209S;

D163N+R181A+G182N+D188N+D199N+D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+D188N+D199N+D205N+M208F+D207N+D209S; and

D163N+R181A+G182N+K185T+A186N+D188N+D199N+D205N+M208F+D207N+D209S.

In a further preferred embodiment the parent alpha-amy- 50 lase is the *B. stearothermophilus* alpha-amylase having the sequence disclosed in SEQ ID NO: 8. In this embodiment preferred substitutions are selected among: D162Q,N, D186N, D203N, M206Y and D207N.

Alterations in the B-domain of SEQ ID NO: 8 where an 55 amino acid is substituted with the corresponding amino acid from the alpha-amylase having the sequence SEQ ID NO: 9 or deletions in case that no corresponding amino acid exists include the alterations: D105N, G108A, G112E, W115V, V116F, D117K, A118V, V119I, N122D, S124N, N127T, 60 Q128K, G132E, T133P, Y134F, Q135E, A138G, D143T, N149D, T150Q, R157N, W158S, Y159E, V164T, W166F, E168A, S169K, R170G, K171E, L172R, S173T, R174G, I175V, Y176F, K177R, F178I, R179A, I181E, G182N, A184K, W187E, E188N, T191D, Y201F, L204I and, E210D. 65

As examples of further alterations known to increase performance of alpha-amylases are: I181\*+G182\*, A184Y/T,

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N191F, N193F, M200X preferably L,I,T,S or A, L204F, M206Y, V212I, S242A,D,E,N or Q, H296Y, G474R and G475R.

As examples or preferred variants according to this embodiment can be mentioned:

D207N+D186N;

D207N+D186N+D162N;

D207N+D186N+D162N+D203N;

D207N+D186N+D162N+D203N+M206Y;

D207N+D186N+D162N+D203N+M206Y+D105N;

D207N+A184K+W187E;

D162N+A184K+D186N+D207N;

D207N+A184K+W187E+D186N;

D207N+A184K+W187E+D186N+D162N;

D207N+A184K+W187E+D186N+D162N+D203N;
 D207N+A184K+W187E+D186N+D162N+D203N+M206Y;
 and

D207N+A184K+W187E+D186N+D162N+D203N+ M206Y+D105N.

Other preferred parent alpha-amylases includes: SEQ ID NO: 2 in WO 2005/001064, *B. licheniformis* alpha-amylase having the sequence SEQ ID NO: 4, *B. amyloliquefaciens* alpha-amylase having the sequence SEQ ID NO: 6, alpha-amylase derived from a strain of the *Bacillus* sp. NCIB 12289, NCIB 12512, NCIB 12513 or DSM 9375, all of which are described in detail in WO 95/26397, and the #707 alpha-amylase described by Tsukamoto et al., 1988, *Biochemical and Biophysical Research Communications*, 151: 25-31 and the alpha-amylase derived from KSM-Ap1378 and described in WO 94/26881.

Nucleotide Sequences

Cloning a DNA Sequence Encoding an Alpha-Amylase

The DNA sequence encoding a parent alpha-amylase may be isolated from any cell or microorganism producing the alpha-amylase in question, using various methods well known in the art. First, a genomic DNA and/or cDNA library should be constructed using chromosomal DNA or messenger RNA from the organism that produces the alpha-amylase to be studied. Then, if the amino acid sequence of the alpha-amylase is known, homologous, labelled oligonucleotide probes may be synthesized and used to identify alpha-amylase-encoding clones from a genomic library prepared from the organism in question. Alternatively, a labelled oligonucleotide probe containing sequences homologous to a known alpha-amylase gene could be used as a probe to identify alpha-amylase-encoding clones, using hybridization and washing conditions of lower stringency.

Yet another method for identifying alpha-amylase-encoding clones would involve inserting fragments of genomic DNA into an expression vector, such as a plasmid, transforming alpha-amylase-negative bacteria with the resulting genomic DNA library, and then plating the transformed bacteria onto agar containing a substrate for alpha-amylase, thereby allowing clones expressing the alpha-amylase to be identified.

Alternatively, the DNA sequence encoding the enzyme may be prepared synthetically by established standard methods, e.g., the phosphoroamidite method described by S. L. Beaucage and M. H. Caruthers (1981) or the method described by Matthes et al. (1984). In the phosphoroamidite method, oligonucleotides are synthesized, e.g., in an automatic DNA synthesizer, purified, annealed, ligated and cloned in appropriate vectors.

Finally, the DNA sequence may be of mixed genomic and synthetic origin, mixed synthetic and cDNA origin or mixed genomic and cDNA origin, prepared by ligating fragments of synthetic, genomic or cDNA origin (as appropriate, the frag-

ments corresponding to various parts of the entire DNA sequence), in accordance with standard techniques. The DNA sequence may also be prepared by polymerase chain reaction (PCR) using specific primers, for instance as described in U.S. Pat. No. 4,683,202 or Saiki et al. (1988).

Site-Directed Mutagenesis

Once an alpha-amylase-encoding DNA sequence has been isolated, and desirable sites for mutation identified, mutations may be introduced using synthetic oligonucleotides. These oligonucleotides contain nucleotide sequences flanking the 10 desired mutation sites; mutant nucleotides are inserted during oligonucleotide synthesis. In a specific method, a singlestranded gap of DNA, bridging the alpha-amylase-encoding sequence, is created in a vector carrying the alpha-amylase gene. Then the synthetic nucleotide, bearing the desired mutation, is annealed to a homologous portion of the singlestranded DNA. The remaining gap is then filled in with DNA polymerase I (Klenow fragment) and the construct is ligated using T4 ligase. A specific example of this method is described in Morinaga et al. (1984). U.S. Pat. No. 4,760,025 20 disclose the introduction of oligonucleotides encoding multiple mutations by performing minor alterations of the cassette. However, an even greater variety of mutations can be introduced at any one time by the Morinaga method, because a multitude of oligonucleotides, of various lengths, can be 25 introduced.

Another method for introducing mutations into alphaamylase-encoding DNA sequences is described in Nelson and Long (1989). It involves the 3-step generation of a PCR fragment containing the desired mutation introduced by using a chemically synthesized DNA strand as one of the primers in the PCR reactions. From the PCR-generated fragment, a DNA fragment carrying the mutation may be isolated by cleavage with restriction endonucleases and reinserted into an expression plasmid.

Homology (Identity)

The homology may be determined as the degree of identity between the two sequences indicating a derivation of the first sequence from the second. The homology may suitably be determined by means of computer programs known in the art such as GAP provided in the GCG program package (described above). Thus, Gap GCGv8 may be used with the default scoring matrix for identity and the following default parameters: GAP creation penalty of 5.0 and GAP extension penalty of 0.3, respectively for nucleic acidic sequence comparison, and GAP creation penalty of 3.0 and GAP extension penalty of 0.1, respectively, for protein sequence comparison. GAP uses the method of Needleman and Wunsch, 1970, *J. Mol. Biol.* 48: 443-453, to make alignments and to calculate the identity.

A structural alignment between Termamyl and a Termamyl-like alpha-amylase may be used to identify equivalent/corresponding positions in other Termamyl-like alphaamylases. One method of obtaining said structural alignment is to use the Pile Up programme from the GCG package using 55 default values of gap penalties, i.e., a gap creation penalty of 3.0 and gap extension penalty of 0.1. Other structural alignment methods include the hydrophobic cluster analysis (Gaboriaud et al., 1987, FEBS Letters 224: 149-155) and reverse threading (Huber et al., 1998, Protein Science 7(1): 60 142-149). Property ii) of the alpha-amylase, i.e., the immunological cross reactivity, may be assayed using an antibody raised against, or reactive with, at least one epitope of the relevant Termamyl-like alpha-amylase. The antibody, which may either be monoclonal or polyclonal, may be produced by methods known in the art, e.g., as described by Hudson et al., Practical Immunology, Third edition (1989), Blackwell Sci18

entific Publications. The immunological cross-reactivity may be determined using assays known in the art, examples of which are Western Blotting or radial immunodiffusion assay, e.g., as described by Hudson et al., 1989. In this respect, immunological cross-reactivity between the alpha-amylases having the amino acid sequences SEQ ID NOS: 2, 4, 6, or 8, respectively, have been found.

Hybridization

The oligonucleotide probe used in the characterization of the Termamyl-like alpha-amylase may suitably be prepared on the basis of the full or partial nucleotide or amino acid sequence of the alpha-amylase in question.

Suitable conditions for testing hybridization involve presoaking in 5×SSC and prehybridizing for 1 hour at ~40° C. in a solution of 20% formamide, 5×Denhardt's solution, 50 mM sodium phosphate, pH 6.8, and 50 mg of denatured sonicated calf thymus DNA, followed by hybridization in the same solution supplemented with 100 mMATP for 18 hours at ~40° C., followed by three times washing of the filter in 2×SSC, 0.2% SDS at 40° C. for 30 minutes (low stringency), preferred at 50° C. (medium stringency), more preferably at 65° C. (high stringency), even more preferably at ~75° C. (very high stringency). More details about the hybridization method can be found in Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, 1989.

In the present context, "derived from" is intended not only to indicate an alpha-amylase produced or producible by a strain of the organism in question, but also an alpha-amylase encoded by a DNA sequence isolated from such strain and produced in a host organism transformed with said DNA sequence. Finally, the term is intended to indicate an alpha-amylase, which is encoded by a DNA sequence of synthetic and/or cDNA origin and which has the identifying characteristics of the alpha-amylase in question. The term is also intended to indicate that the parent alpha-amylase may be a variant of a naturally occurring alpha-amylase, i.e., a variant, which is the result of a modification (insertion, substitution, deletion) of one or more amino acid residues of the naturally occurring alpha-amylase.

Production of Variant Alpha-Amylases

The variant alpha-amylases of the invention may be produced using methods well known in the area. Generally, DNA sequences encoding the parent alpha-amylase is provided and the desired alteration is generated in the nucleic acid sequence using techniques known in the art.

The generated DNA sequence encoding the desired variant alpha-amylase of the invention is provided with suitable regulatory sequences, such as promoter, terminator, activation sites, ribosome binding sites, polyadenylation sites etc. and introduced into a suitable host cell. Finally the host cell comprising said DNA is grown under conditions leading to expression of the variant alpha-amylase according to the invention

All these techniques are known in the art and it is within the skills of the average practitioner within the field to prescribe a suitable method for producing a given variant alpha-amylase of the invention using techniques disclosed in well known text books such as Sambrook et al., Molecular Cloning: A Laboratory Manual, 2nd Ed., Cold Spring Harbor, 1989.

Further teachings regarding preparation of variant alphaamylases can be found in WO 2006/002643, which is incorporated by reference, and the skilled person will appreciate that this teaching also applies to the present invention. Compositions

The present invention also relates to compositions comprising an alpha-amylase variant and at least one additional

enzyme. The additional enzyme(s) may be selected from the group consisting of beta-amylase, cellulase (beta-glucosidase, cellobiohydrolase and endoglucanase), glucoamylase, hemicellulsae (e.g., xylanase), isoamylase, isomerase, lipase, phytase, protease, pullulanase, and/or other enzymes useful 5 in a commercial process in conjunction with an alpha-amylase. The additional enzyme may also be a second alpha-amylase. Such enzymes are known in the art in starch processing, sugar conversion, fermentations for alcohol and other useful end-products, commercial detergents and cleaning aids, stain removal, fabric treatment or desizing, and the like.

Methods of Using the Alpha-Amylase Variants—Industrial Applications

The variants of the present invention possess valuable 15 properties allowing for a variety of industrial applications. In particular, the variants may be used in detergents, in particular laundry detergent compositions and dishwashing detergent compositions, hard surface cleaning compositions, and for desizing textiles, fabrics or garments, production of pulp and 20 paper, beer making, ethanol production, and starch conversion processes.

The alpha-amylase variants may be used for starch processes, in particular starch conversion, especially liquefaction of starch (see, e.g., U.S. Pat. No. 3,912,590, EP 252730 and 25 EP 063909, WO 99/19467, and WO 96/28567, which are all hereby incorporated by reference). Also contemplated are compositions for starch conversion purposes, which may beside the variant of the invention also comprise an AMG, pullulanase, and other alpha-amylases.

Further, the variants are particularly useful in the production of sweeteners and ethanol (see, e.g., U.S. Pat. No. 5,231, 017, which is hereby incorporated by reference), such as fuel, drinking and industrial ethanol, from starch or whole grains.

The variants may also be used for desizing of textiles, 35 fabrics, and garments (see, e.g., WO 95/21247, U.S. Pat. No. 4,643,736, and EP 119920, which are incorporated herein by reference), beer making or brewing, and in pulp and paper production or related processes.

Starch Processing

Native starch consists of microscopic granules, which are insoluble in water at room temperature. When an aqueous starch slurry is heated, the granules swell and eventually burst, dispersing the starch molecules into the solution. During this "gelatinization" process there is a dramatic increase 45 in viscosity. As the solids level is 30-40% in a typical industrial process, the starch has to be thinned or "liquefied" so that it can be suitably processed. This reduction in viscosity is primarily attained by enzymatic degradation in current commercial practice.

Conventional starch-conversion processes, such as liquefaction and saccharification processes are described, e.g., in U.S. Pat. No. 3,912,590, EP 252730 and EP 063909, which are incorporated herein by reference.

In an embodiment, the conversion process degrading starch 55 to lower molecular weight carbohydrate components such as sugars or fat replacers includes a debranching step.

In the case of converting starch into a sugar, the starch is depolymerized. Such a depolymerization process consists of, e.g., a pre-treatment step and two or three consecutive process steps, i.e., a liquefaction process, a saccharification process, and depending on the desired end-product, an optional isomerization process.

When the desired final sugar product is, e.g., high fructose syrup the dextrose syrup may be converted into fructose. 65 After the saccharification process, the pH is increased to a value in the range of 6-8, preferably pH 7.5, and the calcium

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is removed by ion exchange. The dextrose syrup is then converted into high fructose syrup using, e.g., an immobilized glucose isomerase.

Production of Fermentation Products

In general, alcohol production (ethanol) from whole grain can be separated into 4 main steps: milling, liquefaction, saccharification, and fermentation.

The grain is milled in order to open up the structure and allow for further processing. Two processes used are wet or dry milling. In dry milling, the whole kernel is milled and used in the remaining part of the process. Wet milling gives a very good separation of germ and meal (starch granules and protein) and is with a few exceptions applied at locations where there is a parallel production of syrups.

In the liquefaction process the starch granules are solubilized by hydrolysis to maltodextrins mostly of a DP higher than 4. The hydrolysis may be carried out by acid treatment or enzymatically by an alpha-amylase. Acid hydrolysis is used on a limited basis. The raw material can be milled whole grain or a side stream from starch processing.

During a typical enzymatic liquefaction, the long-chained starch is degraded into branched and linear shorter units (maltodextrins) by an alpha-amylase. Enzymatic liquefaction is generally carried out as a three-step hot slurry process. The slurry is heated to between 60-95° C. (e.g., 77-86° C., 80-85° C., or 83-85° C.) and the enzyme(s) is (are) added. The liquefaction process is carried out at 85° C. for 1-2 hours. The pH is generally between 5.5 and 6.2. In order to ensure optimal enzyme stability under these conditions, 1 mM of calcium is added (to provide about 40 ppm free calcium ions). After such treatment, the liquefied starch will have a "dextrose equivalent" (DE) of 10-15.

The slurry is subsequently jet-cooked at between 95-140° C., e.g., 105-125° C., cooled to 60-95° C. and more enzyme(s) is (are) added to obtain the final hydrolysis. The liquefaction process is carried out at pH 4.5-6.5, typically at a pH between 5 and 6. Milled and liquefied grain is also known as mash

Liquefied starch-containing material is saccharified in the presence of saccharifying enzymes such as glucoamylases. The saccharification process may last for 12 hours to 120 hours (e.g., 12 to 90 hours, 12 to 60 hours and 12 to 48 hours).

However, it is common to perform a pre-saccharification step for about 30 minutes to 2 hours (e.g., 30 to 90 minutes) at a temperature of 30 to 65° C., typically around 60° C., which is followed by a complete saccharification during fermentation referred to as simultaneous saccharification and fermentation (SSF). The pH is usually between 4.2-4.8, e.g., 4.5. In a simultaneous saccharification and fermentation (SSF) process, there is no holding stage for saccharification, rather, the yeast and enzymes are added together.

In a typical saccharification process, maltodextrins produced during liquefaction are converted into dextrose by adding a glucoamylase and a debranching enzyme, such as an isoamylase (U.S. Pat. No. 4,335,208) or a pullulanase. The temperature is lowered to 60° C., prior to the addition of a glucoamylase and debranching enzyme. The saccharification process proceeds for 24-72 hours.

Prior to addition of the saccharifying enzymes, the pH is reduced to below 4.5, while maintaining a high temperature (above 95° C.), to inactivate the liquefying alpha-amylase. This process reduces the formation of short oligosaccharide called "panose precursors," which cannot be hydrolyzed properly by the debranching enzyme. Normally, about 0.2-0.5% of the saccharification product is the branched trisaccharide panose (Glc p $\alpha$ 1-6Glc p $\alpha$ 1-4Glc), which cannot be degraded by a pullulanase. If active amylase from the lique-

faction remains present during saccharification (i.e., no denaturing), the amount of panose can be as high as 1-2%, which is highly undesirable since it lowers the saccharification yield significantly.

Fermentable sugars (e.g., dextrins, monosaccharides, particularly glucose) are produced from enzymatic saccharification. These fermentable sugars may be further purified and/or converted to useful sugar products. In addition, the sugars may be used as a fermentation feedstock in a microbial fermentation process for producing end-products, such as alcohol (e.g., ethanol and butanol), organic acids (e.g., succinic acid and lactic acid), sugar alcohols (e.g., glycerol), ascorbic acid intermediates (e.g., gluconate, 2-keto-D-gluconate, 2,5-diketo-D-gluconate, and 2-keto-L-gulonic acid), amino acids (e.g., lysine), proteins (e.g., antibodies and fragment thereof).

In an embodiment, the fermentable sugars obtained during the liquefaction process steps are used to produce alcohol and particularly ethanol. In ethanol production, an SSF process is commonly used wherein the saccharifying enzymes and fermenting organisms (e.g., yeast) are added together and then 20 carried out at a temperature of 30-40° C.

The organism used in fermentation will depend on the desired end-product. Typically, if ethanol is the desired end product yeast will be used as the fermenting organism. In some preferred embodiments, the ethanol-producing micro- 25 organism is a yeast and specifically Saccharomyces such as strains of S. cerevisiae (U.S. Pat. No. 4,316,956). A variety of S. cerevisiae are commercially available and these include but are not limited to FALI (Fleischmann's Yeast), SUPER-START (Alltech), FERMIOL (DSM Specialties), RED 30 STAR (Lesaffre) and Angel alcohol yeast (Angel Yeast Company, China). The amount of starter yeast employed in the methods is an amount effective to produce a commercially significant amount of ethanol in a suitable amount of time, (e.g., to produce at least 10% ethanol from a substrate having 35 between 25-40% DS in less than 72 hours). Yeast cells are generally supplied in amounts of about 10<sup>4</sup> to about 10<sup>12</sup>, and preferably from about 10<sup>7</sup> to about 10<sup>10</sup> viable yeast count per mL of fermentation broth. After yeast is added to the mash, it is typically subjected to fermentation for about 24-96 hours, 40 e.g., 35-60 hours. The temperature is between about 26-34° C., typically at about 32° C., and the pH is from pH 3-6, e.g., around pH 4-5.

The fermentation may include, in addition to a fermenting microorganisms (e.g., yeast), nutrients, and additional 45 enzymes, including phytases. The use of yeast in fermentation is well known in the art.

In further embodiments, use of appropriate fermenting microorganisms, as is known in the art, can result in fermentation end product including, e.g., glycerol, 1,3-propanediol, 50 gluconate, 2-keto-D-gluconate, 2,5-diketo-D-gluconate, 2-keto-L-gulonic acid, succinic acid, lactic acid, amino acids, and derivatives thereof. More specifically when lactic acid is the desired end product, a *Lactobacillus* sp. (*L. casei*) may be used; when glycerol or 1,3-propanediol are the desired end-products *E. coli* may be used; and when 2-keto-D-gluconate, 2,5-diketo-D-gluconate, and 2-keto-L-gulonic acid are the desired end products, *Pantoea citrea* may be used as the fermenting microorganism. The above enumerated list are only examples and one skilled in the art will be aware of a 60 number of fermenting microorganisms that may be used to obtain a desired end product.

Processes for Producing Fermentation Products from Ungelatinized Starch-Containing Material

The invention relates to processes for producing fermentation products from starch-containing material without gelatinization (i.e., without cooking) of the starch-containing mate22

rial. The fermentation product, such as ethanol, can be produced without liquefying the aqueous slurry containing the starch-containing material and water. In one embodiment a process of the invention includes saccharifying (e.g., milled) starch-containing material, e.g., granular starch, below the initial gelatinization temperature, preferably in the presence of alpha-amylase and/or carbohydrate-source generating enzyme(s) to produce sugars that can be fermented into the fermentation product by a suitable fermenting organism. In this embodiment the desired fermentation product, e.g., ethanol, is produced from ungelatinized (i.e., uncooked), preferably milled, cereal grains, such as corn. Accordingly, in the first aspect the invention relates to processes for producing fermentation products from starch-containing material comprising simultaneously saccharifying and fermenting starch-containing material using a carbohydrate-source generating enzyme and a fermenting organism at a temperature below the initial gelatinization temperature of said starchcontaining material. In an embodiment a protease is also present. The protease may be any acid fungal protease or metalloprotease. The fermentation product, e.g., ethanol, may optionally be recovered after fermentation, e.g., by distillation. Typically amylase(s), such as glucoamylase(s) and/ or other carbohydrate-source generating enzymes, and/or alpha-amylase(s), is(are) present during fermentation. Examples of glucoamylases and other carbohydrate-source generating enzymes include raw starch hydrolyzing glucoamylases. Examples of alpha-amylase(s) include acid alpha-amylases such as acid fungal alpha-amylases. Examples of fermenting organisms include yeast, e.g., a strain of Saccharomyces cerevisiae. The term "initial gelatinization temperature" means the lowest temperature at which starch gelatinization commences. In general, starch heated in water begins to gelatinize between about 50° C. and 75° C.; the exact temperature of gelatinization depends on the specific starch and can readily be determined by the skilled artisan. Thus, the initial gelatinization temperature may vary according to the plant species, to the particular variety of the plant species as well as with the growth conditions. In the context of this invention the initial gelatinization temperature of a given starch-containing material may be determined as the temperature at which birefringence is lost in 5% of the starch granules using the method described by Gorinstein and Lii, 1992, Starch/Stärke 44(12): 461-466. Before initiating the process a slurry of starch-containing material, such as granular starch, having 10-55, e.g., 25-45 and 30-40, w/w % dry solids (DS) of starch-containing material may be prepared. The slurry may include water and/or process waters, such as stillage (backset), scrubber water, evaporator condensate or distillate, side-stripper water from distillation, or process water from other fermentation product plants. Because the process of the invention is carried out below the initial gelatinization temperature, and thus no significant viscosity increase takes place, high levels of stillage may be used if desired. In an embodiment the aqueous slurry contains from about 1 to about 70 vol. %, preferably 15-60 vol. %, especially from about 30 to 50 vol. % water and/or process waters, such as stillage (backset), scrubber water, evaporator condensate or distillate, side-stripper water from distillation, or process water from other fermentation product plants, or combinations thereof, or the like. The starch-containing material may be prepared by reducing the particle size, preferably by dry or wet milling, to 0.05 to 3.0 mm, preferably 0.1-0.5 mm. After being subjected to a process of the invention at least 85%, at least 86%, at least 87%, at least 88%, at least 89%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or prefer-

ably at least 99% of the dry solids in the starch-containing material are converted into a soluble starch hydrolyzate. A process in this aspect of the invention is conducted at a temperature below the initial gelatinization temperature, which means that the temperature typically lies in the range between 5 30-75° C., preferably between 45-60° C. In a preferred embodiment the process carried at a temperature from 25° C. to 40° C., such as from 28° C. to 35° C., such as from 30° C. to 34° C., preferably around 32° C. In an embodiment the process is carried out so that the sugar level, such as glucose level, is kept at a low level, such as below 6 w/w %, such as below about 3 w/w %, such as below about 2 w/w %, such as below about 1 w/w %, such as below about 0.5 w/w %, or below 0.25 w/w %, such as below about 0.1 w/w %. Such low levels of sugar can be accomplished by simply employing 15 adjusted quantities of enzyme and fermenting organism. A skilled person in the art can easily determine which doses/ quantities of enzyme and fermenting organism to use. The employed quantities of enzyme and fermenting organism may also be selected to maintain low concentrations of mal- 20 tose in the fermentation broth. For instance, the maltose level may be kept below about 0.5 w/w %, such as below about 0.2 w/w %. The process of the invention may be carried out at a pH from about 3 and 7, preferably from pH 3.5 to 6, or more preferably from pH 4 to 5. In an embodiment fermentation is 25 ongoing for 6 to 120 hours, in particular 24 to 96 hours. Processes for Producing Fermentation Products from Gelati-

nized Starch-Containing Material

In this aspect the invention relates to processes for producing fermentation products, especially ethanol, from starchcontaining material, which process includes a liquefaction step and sequentially or simultaneously performed saccharification and fermentation steps. Consequently, the invention relates to processes for producing fermentation products from starch-containing material comprising the steps of:

- (a) liquefying starch-containing material in the presence of an alpha-amylase variant, or;
- (b) saccharifying the liquefied material obtained in step (a) using a carbohydrate-source generating enzyme;
  - (c) fermenting using a fermenting organism.

In an aspect, a pullulanase such as a family GH57 pullulanase is also used in the liquefaction step. In an embodiment a protease, such as an acid fungal protease or a metalloprotease is added before, during and/or after liquefaction. In an embodiment the metalloprotease is derived from a strain of 45 Thermoascus, e.g., a strain of Thermoascus aurantiacus, especially Thermoascus aurantiacus CGMCC No. 0670. In an embodiment the carbohydrate-source generating enzyme is a glucoamylase derived from a strain of Aspergillus, e.g., Aspergillus niger or Aspergillus awamori, a strain of Talaro- 50 myces, especially Talaromyces emersonii; or a strain of Athelia, especially Athelia rolfsii; a strain of Trametes, e.g., Trametes cingulata; a strain of the genus Pachykytospora, e.g., a strain of *Pachykytospora papyracea*; or a strain of the genus Leucopaxillus, e.g., Leucopaxillus giganteus; or a strain of 55 the genus Peniophora, e.g., a strain of the species Peniophora rufomarginata; or a mixture thereof. Saccharification step (b) and fermentation step (c) may be carried out either sequentially or simultaneously. A pullulanase and/or metalloprotease may be added during saccharification and/or fermenta- 60 tion when the process is carried out as a sequential saccharification and fermentation process and before or during fermentation when steps (b) and (c) are carried out simultaneously (SSF process). The pullulanase and/or metalloprotease may also advantageously be added before liquefaction 65 (pre-liquefaction treatment), i.e., before or during step (a), and/or after liquefaction (post liquefaction treatment), i.e.,

after step (a). The pullulanase is most advantageously added before or during liquefaction, i.e., before or during step (a). The fermentation product, such as especially ethanol, may optionally be recovered after fermentation, e.g., by distillation. The fermenting organism is preferably yeast, preferably a strain of Saccharomyces cerevisiae. In a particular embodiment, the process of the invention further comprises, prior to step (a), the steps of:

- x) reducing the particle size of the starch-containing material, preferably by milling (e.g., using a hammer mill);
- y) forming a slurry comprising the starch-containing material and water.

In an embodiment the particle size is smaller than a #7 screen, e.g., a #6 screen. A #7 screen is usually used in conventional prior art processes. The aqueous slurry may contain from 10-55 w/w % dry solids (DS), e.g., 25-45 and 30-40 w/w % dry solids (DS) of starch-containing material. The slurry is heated to above the gelatinization temperature and an alpha-amylase variant may be added to initiate liquefaction (thinning). The slurry may in an embodiment be jetcooked to further gelatinize the slurry before being subjected to alpha-amylase in step (a). Liquefaction may in an embodiment be carried out as a three-step hot slurry process. The slurry is heated to between 60-95° C., preferably between 70-90° C., such as preferably between 80-85° C. at pH 4-6, preferably 4.5-5.5, and alpha-amylase variant, optionally together with a pullulanase and/or protease, preferably metalloprotease, are added to initiate liquefaction (thinning). In an embodiment the slurry may then be jet-cooked at a temperature between 95-140° C., preferably 100-135° C., such as 105-125° C., for about 1-15 minutes, preferably for about 3-10 minutes, especially around about 5 minutes. The slurry is cooled to 60-95° C. and more alpha-amylase variant and optionally pullulanase variant and/or protease, preferably 35 metalloprotease, is(are) added to finalize hydrolysis (secondary liquefaction). The liquefaction process is usually carried out at pH 4.0-6, in particular at a pH from 4.5 to 5.5. Saccharification step (b) may be carried out using conditions well known in the art. For instance, a full saccharification process may last up to from about 24 to about 72 hours, however, it is common only to do a pre-saccharification of typically 40-90 minutes at a temperature between 30-65° C., typically about 60° C., followed by complete saccharification during fermentation in a simultaneous saccharification and fermentation process (SSF process). Saccharification is typically carried out at temperatures from 20-75° C., preferably from 40-70° C., typically around 60° C., and at a pH between 4 and 5, normally at about pH 4.5. The most widely used process to produce a fermentation product, especially ethanol, is a simultaneous saccharification and fermentation (SSF) process, in which there is no holding stage for the saccharification, meaning that a fermenting organism, such as yeast, and enzyme(s), may be added together. SSF may typically be carried out at a temperature from 25° C. to 40° C., such as from  $28^{\circ}$  C. to  $35^{\circ}$  C., such as from  $30^{\circ}$  C. to  $34^{\circ}$  C., preferably around about 32° C. In an embodiment fermentation is ongoing for 6 to 120 hours, in particular 24 to 96 hours. Beer Making

The alpha-amylase variants may also be used in a beermaking process and similar fermentations; the alpha-amylases will typically be added during the mashing process. The process is substantially similar to the milling, liquefaction, saccharification, and fermentation processes described above.

Starch Slurry Processing with Stillage

Milled starch-containing material is combined with water and recycled thin-stillage resulting in an aqueous slurry. The

slurry can comprise between 15 to 55% ds w/w (e.g., 20 to 50%, 25 to 50%, 25 to 45%, 25 to 40%, 20 to 35% and 30-36% ds). In some embodiments, the recycled thin-stillage (back-set) is in the range of about 10 to 70% v/v (e.g., 10 to 60%, 10 to 50%, 10 to 40%, 10 to 30%, 10 to 20%, 20 to 60%, 20 to 50%, 20 to 40% and also 20 to 30%).

Once the milled starch-containing material is combined with water and backset, the pH is not adjusted in the slurry. Further the pH is not adjusted after the addition of a phytase and optionally an alpha-amylase to the slurry. In an embodiment, the pH of the slurry will be in the range of about pH 4.5 to less than about 6.0 (e.g., pH 4.5 to 5.8, pH 4.5 to 5.6, pH 4.8 to 5.8, pH 5.0 to 5.8, pH 5.0 to 5.4, pH 5.2 to 5.5 and pH 5.2 to 5.9). The pH of the slurry may be between about pH 4.5 and 5.2 depending on the amount of thin stillage added to the 15 slurry and the type of material comprising the thin stillage. For example, the pH of the thin stillage may be between pH 3.8 and pH 4.5.

During ethanol production, acids can be added to lower the pH in the beer well, to reduce the risk of microbial contami- 20 nation prior to distillation.

In some embodiments, a phytase is added to the slurry. In other embodiments, in addition to a phytase, an alpha-amylase is added to the slurry. In some embodiments, a phytase and alpha-amylase are added to the slurry sequentially. In 25 other embodiments, a phytase and alpha-amylase are added simultaneously. In some embodiments, the slurry comprising a phytase and optionally, an alpha-amylase, are incubated (pretreated) for a period of about 5 minutes to about 8 hours (e.g., 5 minutes to 6 hours, 5 minutes to 4 hours, 5 minutes to 2 hours, and 15 minutes to 4 hours). In other embodiments, the slurry is incubated at a temperature in the range of about 40 to 115° C. (e.g., 45 to 80° C., 50 to 70° C., 50 to 75° C., 60 to 110° C., 60 to 95° C., 70 to 110° C., 70 to 85° C. and 77 to 86° C.).

In other embodiments, the slurry is incubated at a temperature of about 0 to about 30° C. (e.g., 0 to 25° C., 0 to 20° C., 0 to 15° C., 0 to 10° C. and 0 to 5° C.) below the starch gelatinization temperature of the starch-containing material. In some embodiments, the temperature is below about  $68^{\circ}$  C., 40 below about  $65^{\circ}$  C., below about  $62^{\circ}$  C., below about  $60^{\circ}$  C. and below about  $55^{\circ}$  C. In some embodiments, the temperature is above about  $45^{\circ}$  C., above about  $50^{\circ}$  C., above about  $55^{\circ}$  C. and above about  $60^{\circ}$  C. In some embodiments, the incubation of the slurry comprising a phytase and an alphaamylase at a temperature below the starch gelatinization temperature is referred to as a primary  $(1^{\circ})$  liquefaction.

In one embodiment, the milled starch-containing material is corn or milo. The slurry comprises 25 to 40% DS, the pH is in the range of 4.8 to 5.2, and the slurry is incubated with a 50 phytase and optionally an alpha-amylase for 5 minutes to 2 hours, at a temperature range of 60 to 75° C.

Currently, it is believed that commercially-available or microbial alpha-amylases used in the liquefaction process are generally not stable enough to produce liquefied starch substrate from a dry mill process using whole ground grain at a temperature above about 80° C. at a pH level that is less than pH 5.6. The stability of many commercially available alphaamylases is reduced at a pH of less than about 4.0.

In a further liquefaction step, the incubated or pretreated 60 starch-containing material is exposed to an increase in temperature such as about 0 to about 45° C. above the starch gelatinization temperature of the starch-containing material (e.g., 70° C. to 120° C., 70° C. to 110° C., and 70° C. to 90° C.) for a period of time of about 2 minutes to about 6 hours (e.g., 2 minutes to 4 hrs, 90 minutes, 140 minutes and 90 to 140 minutes) at a pH of about 4.0 to 5.5 more preferably

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between 1 hour to 2 hours. The temperature can be increased by a conventional high temperature jet cooking system for a short period of time, for example, for 1 to 15 minutes. Then the starch maybe further hydrolyzed at a temperature ranging from about 75° C. to 95° C. (e.g., 80° C. to 90° C. and 80° C. to 85° C.) for a period of about 15 to 150 minutes (e.g., 30 to 120 minutes). In a preferred embodiment, the pH is not adjusted during these process steps and the pH of the liquefied mash is in the range of about pH 4.0 to pH 5.8 (e.g., pH 4.5 to 5.8, pH 4.8 to 5.4, and pH 5.0 to 5.2). In some embodiments, a second dose of thermostable alpha-amylase is added to the secondary liquefaction step, but in other embodiments there is no additional dosage of alpha-amylase.

The incubation and liquefaction steps may be followed by saccharification and fermentation steps well known in the art. Distillation

Optionally, following fermentation, an alcohol (e.g., ethanol) may be extracted by, for example, distillation and optionally followed by one or more process steps.

In some embodiments, the yield of ethanol produced by the methods provided herein is at least 8%, at least 10%, at least 12%, at least 14%, at least 15%, at least 16%, at least 17% and at least 18% (v/v) and at least 23% v/v. The ethanol obtained according to the process provided herein may be used as, for example, fuel ethanol, drinking ethanol, i.e., potable neutral spirits, or industrial ethanol.

By-Products

Left over from the fermentation is the grain, which is typically used for animal feed either in liquid or dried form. In further embodiments, the end product may include the fermentation co-products such as distiller's dried grains (DDG) and distiller's dried grain plus solubles (DDGS), which may be used, for example, as an animal feed.

Further details on how to carry out liquefaction, sacchari-35 fication, fermentation, distillation, and recovery of ethanol are well known to the skilled person.

According to the process provided herein, the saccharification and fermentation may be carried out simultaneously or separately.

O Pulp and Paper Production

The alpha-amylase variants may also be used in the production of lignocellulosic materials, such as pulp, paper and cardboard, from starch reinforced waste paper and cardboard, especially where re-pulping occurs at pH above 7 and where amylases facilitate the disintegration of the waste material through degradation of the reinforcing starch. The alpha-amylase variants are especially useful in a process for producing a papermaking pulp from starch-coated printed-paper. The process may be performed as described in WO 95/14807, comprising the following steps:

- a) disintegrating the paper to produce a pulp,
- b) treating with a starch-degrading enzyme before, during or after step a), and
- c) separating ink particles from the pulp after steps a) and b).

The alpha-amylase variants may also be useful in modifying starch where enzymatically modified starch is used in papermaking together with alkaline fillers such as calcium carbonate, kaolin and clays. With the alpha-amylase variants it is possible to modify the starch in the presence of the filler thus allowing for a simpler integrated process.

Desizing of Textiles, Fabrics and Garments

The alpha-amylase variants may also be very useful in textile, fabric or garment desizing. In the textile processing industry, alpha-amylases are traditionally used as auxiliaries in the desizing process to facilitate the removal of starch-containing size, which has served as a protective coating on

weft yarns during weaving. Complete removal of the size coating after weaving is important to ensure optimum results in the subsequent processes, in which the fabric is scoured, bleached and dyed. Enzymatic starch breakdown is preferred because it does not involve any harmful effect on the fiber 5 material. In order to reduce processing cost and increase mill throughput, the desizing process is sometimes combined with the scouring and bleaching steps. In such cases, non-enzymatic auxiliaries such as alkali or oxidation agents are typically used to break down the starch, because traditional alphaamylases are not very compatible with high pH levels and bleaching agents. The non-enzymatic breakdown of the starch size leads to some fiber damage because of the rather aggressive chemicals used. Accordingly, it would be desirable to use the alpha-amylase variants as they have an improved performance in alkaline solutions. The alpha-amylase variants may be used alone or in combination with a cellulase when desizing cellulose-containing fabric or textile.

Desizing and bleaching processes are well known in the art. For instance, such processes are described in, e.g., WO 20 95/21247, U.S. Pat. No. 4,643,736, EP 119920, which are hereby incorporated by reference.

Cleaning Processes and Detergent Compositions

The alpha-amylase variants may be added as a component of a detergent composition for various cleaning or washing 25 processes, including laundry and dishwashing. For example, the variants may be used in the detergent compositions described in WO 96/23874 and WO 97/07202.

The alpha-amylase variants may be incorporated in detergents at conventionally employed concentrations. For 30 example, a variant of the invention may be incorporated in an amount corresponding to 0.00001-10 mg (calculated as pure, active enzyme protein) of alpha-amylase per liter of wash/ dishwash liquor using conventional dosing levels of deter-

The detergent composition may for example be formulated as a hand or machine laundry detergent composition, including a laundry additive composition suitable for pretreatment of stained fabrics and a rinse added fabric softener composition or be formulated as a detergent composition for use in 40 general household hard surface cleaning operations, or be formulated for hand or machine dishwashing operations.

The detergent composition may further comprise one or more other enzymes, such as a lipase, peroxidase, protease, another amylolytic enzyme, e.g., another alpha-amylase, glu-45 coamylase, maltogenic amylase, CGTase, cellulase, mannanase (such as Mannaway<sup>TM</sup> from Novozymes, Denmark)), pectinase, pectin lyase, cutinase, and/or laccase.

In general the properties of the chosen enzyme(s) should be compatible with the selected detergent (i.e., pH-optimum, 50 compatibility with other enzymatic and non-enzymatic ingredients, etc.), and the enzyme(s) should be present in effective

The detergent enzyme(s) may be included in a detergent more enzymes, or by adding a combined additive comprising all of these enzymes. A detergent additive, e.g., a separate additive or a combined additive, can be formulated, e.g., granulate, a liquid, a slurry, etc. Preferred detergent additive formulations are granulates, in particular non-dusting granu- 60 lates, liquids, in particular stabilized liquids, or slurries.

Non-dusting granulates may be produced, e.g., as disclosed in U.S. Pat. Nos. 4,106,991 and 4,661,452 and may optionally be coated by methods known in the art. Examples of waxy coating materials are poly(ethylene oxide) products 65 (polyethyleneglycol, PEG) with mean molar weights of 1000 to 20000; ethoxylated nonyl-phenols having from 16 to 50

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ethylene oxide units; ethoxylated fatty alcohols in which the alcohol contains from 12 to 20 carbon atoms and in which there are 15 to 80 ethylene oxide units; fatty alcohols, fatty acids; and mono- and di- and triglycerides of fatty acids. Examples of film-forming coating materials suitable for application by fluid bed techniques are given in GB 1483591. Liquid enzyme preparations may, for instance, be stabilized by adding a polyol such as propylene glycol, a sugar or sugar alcohol, lactic acid or boric acid according to established methods. Protected enzymes may be prepared according to the method disclosed in EP 238216.

The detergent composition may be in any convenient form, e.g., a bar, a tablet, a powder, a granule, a paste or a liquid. A liquid detergent may be aqueous, typically containing up to about 70% water and 0 to about 30% organic solvent, or non-aqueous.

The detergent composition comprises one or more surfactants, which may be non-ionic including semi-polar and/or anionic and/or cationic and/or zwitterionic. The surfactants are typically present at a level of from about 0.1% to 60% by

When included therein the detergent will usually contain from about 1% to about 40% of an anionic surfactant such as linear alkylbenzenesulfonate, alpha-olefinsulfonate, alkyl sulfate (fatty alcohol sulfate), alcohol ethoxysulfate, secondary alkanesulfonate, alpha-sulfo fatty acid methyl ester, alkyl- or alkenylsuccinic acid or soap.

When included therein the detergent will usually contain from about 0.2% to about 40% of a non-ionic surfactant such as alcohol ethoxylate, nonyl-phenol ethoxylate, alkylpolyglycoside, alkyldimethylamine-oxide, ethoxylated fatty acid monoethanol-amide, fatty acid monoethanolamide, polyhydroxy alkyl fatty acid amide, or N-acyl N-alkyl derivatives of glucosamine ("glucamides").

The detergent may contain 0 to about 65% of a detergent builder or complexing agent such as zeolite, diphosphate, triphosphate, phosphonate, carbonate, citrate, nitrilotriacetic acid, ethylenediaminetetraacetic acid, diethylenetriaminepentaacetic acid, alkyl- or alkenylsuccinic acid, soluble silicates or layered silicates (e.g., SKS-6 from Hoechst).

The detergent may comprise one or more polymers. Examples are carboxymethylcellulose, poly(vinyl-pyrrolidone), poly(ethylene glycol), poly(vinyl alcohol), poly(vinylpyridine-N-oxide), poly(vinylimidazole), polycarboxylates such as polyacrylates, maleiclacrylic acid copolymers and lauryl methacrylate/acrylic acid co-polymers.

The detergent may contain a bleaching system, which may comprise a H<sub>2</sub>O<sub>2</sub> source such as perborate or percarbonate which may be combined with a peracid-forming bleach activator such as tetraacetylethylenediamine or nonanoyloxyben-zenesul-fonate. Alternatively, the bleaching system may comprise peroxy acids of, e.g., the amide, imide, or sulfone

The enzyme(s) of the detergent composition may be stabicomposition by adding separate additives containing one or 55 lized using conventional stabilizing agents, e.g., a polyol such as propylene glycol or glycerol, a sugar or sugar alcohol, lactic acid, boric acid, or a boric acid derivative, e.g., an aromatic borate ester, or a phenyl boronic acid derivative such as 4-formylphenyl boronic acid, and the composition may be formulated as described in, e.g., WO 92/19708 and WO

> The detergent may also contain other conventional detergent ingredients such as, e.g., fabric conditioners including clays, foam boosters, suds suppressors, anti-corrosion agents, soil-suspending agents, anti-soil re-deposition agents, dyes, bactericides, optical brighteners, hydrotropes, tarnish inhibitors, or perfumes.

The detergent compositions may comprise any enzyme in an amount corresponding to 0.01-100 mg of enzyme protein per liter of wash liquor, preferably 0.055 mg of enzyme protein per liter of wash liquor, in particular 0.1-1 mg of enzyme protein per liter of wash liquor. One or more of the variant 5 enzymes described herein may additionally be incorporated in the detergent formulations disclosed in WO 97/07202, which is hereby incorporated as reference.

This disclosure includes further detail in the following examples, which are not in any way intended to limit the scope of what is claimed. The following examples are thus offered to illustrate, but not to limit what is claimed.

#### **EXAMPLES**

Materials

Enzymes

SP722: SEQ ID NO: 10, available from Novozymes, and disclosed in WO 95/26397.

AA560: SEQ ID NO: 2; disclosed in WO 00/60060 and available from Novozymes A/S; disclosed in Danish patent application no. PA 1999 00490; deposited on Jan. 25, 1999 at DSMZ and assigned the DSMZ no. 12649.

Bacillus subtilis SHA273: see WO 95/10603

Methods

General Molecular Biology Methods

Unless otherwise mentioned the DNA manipulations and transformations were performed using standard methods of molecular biology (Sambrook et al. (1989); Ausubel et al. (1995); Harwood and Cutting (1990)).

Fermentation of Alpha-Amylases and Variants

Fermentation may be performed by methods well known in the art or as follows.

A *B. subtilis* strain harboring the relevant expression plasmid is streaked on a LB-agar plate with a relevant antibiotic, and grown overnight at 37° C.

The colonies are transferred to 100 ml BPX media supplemented with a relevant antibiotic (for instance 10 mg/l chloroamphinicol) in a 500 ml shaking flask.

Composition of BPX medium:

Potato starch	100 g/l
Barley flour	50 g/l
BAN 5000 SKB	0.1 g/l
Sodium caseinate	10 g/l
Soy Bean Meal	20 g/l
Na <sub>2</sub> HPO <sub>4</sub> , 12 H <sub>2</sub> O	9 g/l
Pluronic TM	0.1 g/l
	-

The culture is shaken at 37° C. at 270 rpm for 4 to 5 days.

Cells and cell debris are removed from the fermentation broth by centrifugation at 4500 rpm in 20-25 minutes. Afterwards the supernatant is filtered to obtain a completely clear solution. The filtrate is concentrated and washed on an UF-filter (10000 cut off membrane) and the buffer is changed to 20 mM acetate pH 5.5. The UF-filtrate is applied on an S-sepharose F.F. and elution is carried out by step elution with 60 0.2 M NaCl in the same buffer. The eluate is dialysed against 10 mM Tris, pH 9.0 and applied on a Q-sepharose F.F. and eluted with a linear gradient from 0-0.3 M NaCl over 6 column volumes. The fractions, which contain the activity (measured by the Phadebas assay) are pooled, pH was adjusted to 65 pH 7.5 and remaining color was removed by a treatment with 0.5% w/vol. active coal in 5 minutes.

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Determination of Residual Activity at pH 4.0 Reagents etc.

Buffer: 50 mM Citrate, pH 4, 0.05% Triton-X100

Prepare 500 mM stock solution.

Stability buffer: 50 mM Carbonate buffer (NaHCO<sub>3</sub>), pH 8, 1 mM CaCl<sub>2</sub>, 0.05% Triton-X100
 Sample Preparation

Samples are centrifuged at 20,000 rpm for 2 minutes. If necessary the samples may be diluted in a stability buffer. Incubation

100 microliters prepared sample

Ad 1000 microliters buffer

Incubate at  $35^{\circ}$  C. and withdraw 20 microliters aliquots in a 200 microliters cold stability buffer after 0, 20 and 48 hours. These can be stored on ice for later activity determination.

Measure the residual activity using the Phabedas assay, see protocol below.

Measurement of the Calcium- and pH-Dependent Stability Normally industrial liquefaction processes runs using pH 6.0-6.2 as liquefaction pH and an addition of 40 ppm free calcium in order to improve the stability at 95° C.-105° C. Some of the herein proposed substitutions have been made in order to improve the stability at

5 1. pH lower than pH 6.2 and/or

2. free calcium levels lower than 40 ppm.

Two different methods can be used to measure the alterations in stability obtained by the different substitutions in the alpha-amylase in question:

Method 1. One assay which measures the stability at reduced pH, pH 5.0, in the presence of 5 ppm free calcium.

10 micro g of the variant are incubated under the following conditions: A 0.1 M acetate solution, pH adjusted to pH 5.0, containing 5 ppm calcium and 5% w/w common corn starch (free of calcium). Incubation is made in a water bath at  $95^{\circ}\,\mathrm{C}$ . for 30 minutes.

Method 2. One assay, which measure the stability in the absence of free calcium and where the pH is maintained at pH 6.0. This assay measures the decrease in calcium sensitivity: 10 micro g of the variant were incubated under the following conditions: A 0.1 M acetate solution, pH adjusted to pH 6.0, containing 5% w/w common corn starch (free of calcium). Incubation was made in a water bath at 95° C. for 30 minutes. Assays for Alpha-Amylase Activity

45 1. Phadebas Assay

Alpha-amylase activity is determined by a method employing Phadebas® tablets as substrate. Phadebas tablets (Phadebas® Amylase Test, supplied by Pharmacia Diagnostic) contain a cross-linked insoluble blue-colored starch polymer, which has been mixed with bovine serum albumin and a buffer substance and tabletted.

For every single measurement one tablet is suspended in a tube containing 5 ml 50 mM Britton-Robinson buffer (50 mM acetic acid, 50 mM phosphoric acid, 50 mM boric acid, 0.1 mM CaCl $_2$ , pH adjusted to the value of interest with NaOH). The test is performed in a water bath at the temperature of interest. The alpha-amylase to be tested is diluted in x ml of 50 mM Britton-Robinson buffer. 1 ml of this alpha-amylase solution is added to the 5 ml 50 mM Britton-Robinson buffer. The starch is hydrolyzed by the alpha-amylase giving soluble blue fragments. The absorbance of the resulting blue solution, measured spectrophotometrically at 620 nm, is a function of the alpha-amylase activity.

It is important that the measured 620 nm absorbance after 10 or 15 minutes of incubation (testing time) is in the range of 0.2 to 2.0 absorbance units at 620 nm. In this absorbance range there is linearity between activity and absorbance

32 Example 1

(Lambert-Beer law). The dilution of the enzyme must therefore be adjusted to fit this criterion. Under a specified set of conditions (temp., pH, reaction time, buffer conditions) 1 mg of a given alpha-amylase will hydrolyze a certain amount of substrate and a blue colour will be produced. The colour intensity is measured at 620 nm. The measured absorbance is directly proportional to the specific activity (activity/mg of pure alpha-amylase protein) of the alpha-amylase in question under the given set of conditions.

#### 2. Alternative Method

Alpha-amylase activity is determined by a method employing the PNP-G7 substrate. PNP-G7 which is a abbreviation for p-nitrophenyl-alpha,D-maltoheptaoside is a blocked oligosaccharide which can be cleaved by an endoamylase. Following the cleavage, the alpha-Glucosidase included in the kit digest the substrate to liberate a free PNP molecule which has a yellow colour and thus can be measured by visible spectophometry at  $\lambda$ =405 nm (400-420 nm). Kits containing PNP-G7 substrate and alpha-Glucosidase is 20 manufactured by Boehringer-Mannheim (cat. no. 1054635).

To prepare the substrate one bottle of substrate (BM 1442309) is added to 5 ml buffer (BM1442309). To prepare the alpha-glucosidase one bottle of alpha-glucosidase (BM 1462309) is added to 45 ml buffer (BM1442309). The work- 25 ing solution is made by mixing 5 ml alpha-Glucosidase solution with 1 ml substrate.

The assay is performed by transforming 20 microliters enzyme solution to a 96 well microtitre plate and incubating at  $25^{\circ}$  C. 200 microliters working solution,  $25^{\circ}$  C. is added. 30 The solution is mixed and pre-incubated 1 minute and absorption is measured every 15 sec. over 3 minutes at OD 405 nm.

The slope of the time dependent absorption-curve is directly proportional to the specific activity (activity per mg enzyme) of the alpha-amylase in question under the given set 35 of conditions.

# 3. Enzchek® Amylase Activity Assay

Alpha-amylase activity may also be determined by a method employing the EnzChek® substrate. The substrate in the EnzChek® Ultra Amylase Assay Kit (E33651, Invitrogen, 40 La Jolla, Calif., USA) is a corn starch derivative, DQTM starch, which is corn starch labeled with BODIPY® FL dye to such a degree that fluorescence is quenched.

One vial containing approx. 1 mg lyophilized substrate is dissolved in 100 microliters of 50 mM sodium acetate (pH 45 4.0). The vial is vortexed for 20 seconds and left at room temperature, in the dark, with occasional mixing until dissolved. Then 900 microliters of 100 mM acetate, 0.01% (w/v) TRITON® X100, 0.12 mM CaCl<sub>2</sub>, pH 5.5 is added, vortexed thoroughly and stored at room temperature, in the dark until 50 ready to use. The substrate working solution is prepared by diluting 10-fold in residual activity buffer (100 mM acetate, 0.01% (w/v) TRITON® X100, 0.12 mM CaCl<sub>2</sub>, pH 5.5) giving a substrate concentration of 100 micrograms/ml. Immediately after incubation the enzyme is diluted to a concentration of 20 ng enzyme protein/mL in 100 mM acetate, 0.01% (W/v) TRITON® X100, 0.12 mM CaCl<sub>2</sub>, pH 5.5.

For the assay, 25 microliters of the substrate working solution is mixed for 10 second with 25 microliters of the diluted enzyme in a black 384 well microtiter plate. The fluorescence intensity is measured (excitation: 485 nm, emission: 555 nm) once every minute for 15 minutes in each well at 25° C. and the  $V_{max}$  is calculated as the slope of the plot of fluorescence intensity against time. The plot should be linear and the residual activity assay has been adjusted so that the diluted reference enzyme solution is within the linear range of the activity assay.

#### Preparation of Variants

Using the parent alpha-amylase AA560+delta(D183+G184)+N195F+R118K+R320K+R458K (disclosed in WO 01/66712) following variants were constructed:

- 1. D163Q+D188N+M208F+D209S+K242P+S244W;
- 10 2. D163N+R181A+G182A+K185T+G186N+D188N+D205N+M208F+D209S+V238I+K242P+S244W;
  - 3. N128W+D163N+R181A+G182N+K185T+G186N+ D188N+D205N+M208F+D209S+K242P+S244W;
- 4. D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+N409D+ D432N+A434P;
  - 5. D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W

#### Example 2

#### Measurement of Residual Activity at pH 4.0

The residual activity at pH 4.0 was measured for the parent amylase and 5 variants prepared in example 1. All measurements were done as double determinations. The results are expressed in percent of the initial activity.

The following results were obtained demonstrating the improved stability of the variants of the invention at pH 4.0:

0 hours	20 hours	48 hours
100%	35%	25%
100%	65%	34%
100%	80%	70%
100%	87%	85%
100%	88%	84%
100%	97%	96%
	100% 100% 100% 100%	100% 35% 100% 65% 100% 80% 100% 87% 100% 88%

Example 3

#### Residual Activity After Incubation with Strong Chelators

The parent alpha-amylase and variant 5 described in example 1 was incubated in the presence of strong chelators.

For incubation with chelators following mixtures were prepared:

 $100~\mathrm{microliters}~250~\mathrm{mM}$  EDTA or  $100~\mathrm{microliters}~10\%$  DTPA

100 microliters enzyme preparation

Ad 1000 microliters with buffer.

Samples were incubated at  $35^{\circ}$  C. for 18 hours and the activity was determined using the PNP-G7 method described above.

	Parent	Variant 5
DTPA	6%	42%
EDTA	15%	79%

The results clearly show that the variant of the invention is considerably more resistant to the presence of strong chelators than the parent alpha-amylase.

#### Example 4

#### Measurement of Residual Activity at pH 3.0

Variant 5 and the parent alpha-amylase described in example 1 were incubated at pH 3.0 at  $35^{\circ}$  C. for 18 hours and  $^{10}$  the residual activity was determined using the PNP-G7 assay described above.

	Parent	Variant 5	
pH 3	1%	79%	

The results show the the variant of the invention also 0 has improved stability at pH 3.compared with the parent alpha- 20 amylase.

The invention described and claimed herein is not to be limited in scope by the specific embodiments herein disclosed, since these embodiments are intended as illustrations of several aspects of the invention. Any equivalent embodiments are intended to be within the scope of this invention. Indeed, various modifications of the invention in addition to those shown and described herein will become apparent to those skilled in the art from the foregoing description. Such modifications are also intended to fall within the scope of the appended claims. In the case of conflict, the present disclosure including definitions will control.

Various references are cited herein, the disclosures of which are incorporated by reference in their entireties.

The invention is further defined in the following para- 35 graphs:

Paragraph 1. An isolated variant alpha-amylase, comprising two or more alterations at positions corresponding to positions 163, 188, 205, 208, and 209 of the mature polypeptide of SEQ ID NO: 2, wherein

- (a) the variant has a sequence identity to any of SEQ ID NOS: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, and 17 of at least 70% and less than 100%.
- (b) each alteration is independently a substitution, deletion or insertion; and
- (c) the variant has alpha-amylase activity.

Paragraph 2. The variant of paragraph 1, which has at least 80% sequence identity to sequence identity to any of SEQ ID NOS: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, and 17.

Paragraph 3. The variant of paragraph 1, which at least 90% 50 sequence identity to sequence identity to any of SEQ ID NOS: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, and 17.

Paragraph 4. The variant of paragraph 1, which at least 95% sequence identity to sequence identity to any of SEQ ID NOS: 2, 4, 6, 8, 10, 11, 12, 13, 14, 15, 16, and 17.

Paragraph 5. The variant of any of paragraphs 1-4, which wherein the alterations at positions 163, 188, 205, 208, and 209 are substitutions.

Paragraph 6. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to 60 positions 163 and 188.

Paragraph 7. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163 and 205.

Paragraph 8. The variant of any of paragraphs 1-4, which 65 comprises a substitution at the positions corresponding to positions 163 and 208.

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Paragraph 9. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163 and 209.

Paragraph 10. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188 and 205.

Paragraph 11. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188 and 208.

Paragraph 12. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188 and 209.

Paragraph 13. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 205 and 208.

Paragraph 14. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 205 and 209.

Paragraph 15. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 208 and 209.

Paragraph 16. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, and 205.

Paragraph 17. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, and 208.

Paragraph 18. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, and 209.

Paragraph 19. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188, 205, and 208.

5 Paragraph 20. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188, 205, and 209.

Paragraph 21. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 205, 208, and 209.

Paragraph 22. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, 205, and 208.

Paragraph 23. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, 205, and 209.

Paragraph 24. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 188, 205, 208, and 209.

Paragraph 25. The variant of any of paragraphs 1-4, which comprises a substitution at the positions corresponding to positions 163, 188, 205, 208, and 209.

Paragraph 26. The variant of any of paragraphs 1-25, wherein the alterations are selected among: X163Q,N, X188N, X205N, X208Y and X209N,S.

Paragraph 27. The variant of any of paragraphs 1-26, further comprising one or more alterations selected from the group consisting of a deletion at a position corresponding to positions 183 and 184 and a substitution at a position corresponding to the positions selected from the group consisting of 186, 193, 195, 202, 206, 214, 244, 452, 474 and 475

Paragraph 28. The variant of any of paragraphs 1-27, which further comprises one or more alterations selected from the group of X181\*+X182\*, X182\*+X183\*, X183\*+X184\*, X185K, X167W, X202L/I/T, X203Y, X167W+X168E+X169E+X170R, X51T+X109G+X203Y, X109G+X203Y, X189W, X189W+x190E+x193T, X190E, X193T, X303K,

X303K+x305R+x306D+X409N+X432N+X434D, X305R, X306D, X409N, X432N, X434D.

Paragraph 29. The variant of any of paragraphs 1-28, which further comprises one or more alterations selected from the group consisting of A113E, N116V, V117F, L118K, A119V, 5 V120I, N123D, N126D, N128T, Q129K, G133E, D134P, Y135F, T136E, A139G, D144T, N150D, T151Q, D154S, R158N, W159S, Y160E, V165T, W167F, Q169A, S170K, R171G, Q172\*, F173E, Q174R, N175T, R176G, 1177V, Y178F, K179R, F180I, R181A, D183E, G184N, A186K, 10 W189E, E190N, S193T/D, N195F, Y203F,V206I, E212D, V214R, and N215R.

Paragraph 30. The variant of any of paragraphs 1-29, which further comprises one or more alterations selected from the group consisting of D183\*+G184\*, G186A,Y,T, T193F, 15 N195F, M202L,I,T,S,A, I206F,Y, V214I, S244A,D,E,N,Q, W, T452HY, G474R, G475R.

Paragraph 31. The variant of any of paragraphs 1-30 selected from the group consisting of:

D188N+D209S:

D163N+D188N+D209S;

D163N+D188N+D205N+D209S;

D163N+D188N+D205N+M208F+D209S;

D207N+D209S:

D163N+D207N+D209S;

D163N+D188N+D207N+D209S;

D163N+D188N+D199N+D207N+D209S;

D163N+D188N+D199N+D205N+D207N+D209S;

D163N+D188N+D199N+D205N+M208F+D207N+D209S;

D163N+R181A+G182N+G186N+D188N+D205N+D209S; D163N+R181A+G182N+K185T+G186N+D188N+ D205N+D209S;

D163N+R181A+G182N+K185T+G186N+D188N+D205N+M208F+D209S;

D163N+R181A+G182N+D188N+D199N+D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+D188N+D199N+ D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+G186N+D188N+

D199N+D205N+M208F+D207N+D209S; D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+N409D+ D432N+A434P;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+H408W+ N409D+D432N+A434P;

N128W+D163N+R181A+G182N+K185T+G186N+ D188N+D205N+M208F+D209S+K242P+S244W; D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+V238I+K242P+S244W; and

D163Q+D188N+M208F+D209S+K242P+S244W.

Paragraph 32. The variant of any of paragraphs 1-31 selected 55 from the group consisting of:

D188N+D209S

D163N+D188N+D209S

D163N+D188N+D205N+D209S

D163N+D188N+D205N+M208F+D209S

D207N+D209S

D163N+D207N+D209S

D163N+D188N+D207N+D209S

D163N+D188N+D199N+D207N+D209S

D163N+D188N+D199N+D205N+D207N+D209S

D163N+D188N+D199N+D205N+M208F+D207N+D209S

D163N+R181A+G182N+G186N+D188N+D205N+D209S

36

D163N+R181A+G182N+K185T+A186N+D188N+D205N+D209S

D163N+R181A+G182N+K185T+A186N+D188N+ D205N+M208F+D209S

5 D163N+R181A+G182N+D188N+D199N+D205N+ M208F+D207N+D209S

D163N+R181A+G182N+K185T+D188N+D199N+ D205N+M208F+D207N+D209S

D163N+R181A+G182N+K185T+A186N+D188N+

D199N+D205N+M208F+D207N+D209S.

Paragraph 33. The variant of any of paragraphs 1-32 selected from the group consisting of: D207N+D186N

D207N+D186N+D162N

- D207N D100N D102N

5 D207N+D186N+D162N+D203N D207N+D186N+D162N+D203N+M206Y

D207N+D186N+D162N+D203N+M206Y+D105N

D207N+A184K+T187E

D207N+A184K+T187E+D186N

20 D207N+A184K+T187E+D186N+D162N D207N+A184K+T187E+D186N+D162N+D203N D207N+A184K+T187E+D186N+D162N+D203N+M206Y D207N+A184K+T187E+D186N+D162N+D203N+ M206Y+D105N.

25 Paragraph 34. A detergent composition comprising a variant of any of paragraphs 1-33 and a surfactant.

Paragraph 35. A composition comprising a variant of any of paragraphs 1-33 and one or more enzymes selected from the group consisting of beta-amylase, cellulase (beta-glucosi-

dase, cellobiohydrolase, and endoglucanase) glucoamylase, hemicellulase (e.g., xylanase), isoamylase, isomerase, lipase, phytase, protease, and pullulanase.

Paragraph 36. Use of a variant of any of paragraphs 1-33 for washing and/or dishwashing.

35 Paragraph 37. Use of a variant of any of paragraphs 1-33 for desizing a textile.

Paragraph 38. Use of a variant of any of paragraphs 1-33 for producing a baked product.

Paragraph 39. Use of a variant of any of paragraphs 1-33 for liquefying a starch-containing material.

Paragraph 40. A method of producing liquefied starch, comprising liquefying a starch-containing material with a variant of any of paragraphs 1-33

Paragraph 41. A process of producing a fermentation product, 45 comprising

(a) liquefying a starch-containing material with a variant of any of paragraphs 1-33 to produce a liquefied mash;

(b) saccharifying the liquefied mash to produce fermentable sugars; and

(c) fermenting the fermentable sugars in the presence of a fermenting organism.

Paragraph 42. The process of paragraph 41 wherein the starch-containing material is liquefied with the variant and a pullulanase, e.g., a GH57 pullulanase.

5 Paragraph 43. The process of paragraph 42 wherein the pullulanase is obtained from a strain of *Thermococcus*, including *Thermococcus* sp. AM4, *Thermococcus* sp. HJ21, *Thermococcus barophilus*, *Thermococcus gammatolerans*, *Thermococcus hydrothermalis*; *Thermococcus kodakarensis*, *Ther-*

60 mococcus litoralis, and Thermococcus onnurineus; or from a strain of Pyrococcus, such as Pyrococcus abyssi and Pyrococcus furiosus.

Paragraph 44. The process of any of paragraphs 41-43 further comprising adding a protease, such as an acid fungal protease or a metalloprotease before, during and/or after liquefaction.

Paragraph 45. The process of paragraph 44, wherein the metalloprotease is obtained from a strain of *Thermoascus*, prefator

erably a strain of *Thermoascus aurantiacus*, especially *Thermoascus aurantiacus* CGMCC No. 0670.

Paragraph 46. A process of producing a fermentation product, comprising contacting a starch substrate with a variant of any of paragraphs 1-33, a glucoamylase, and a fermenting organism.

Paragraph 47. The process of any of paragraphs 41-46, wherein the fermentation product is selected from the group consisting of alcohol (e.g., ethanol and butanol), organic acids (e.g., succinic acid and lactic acid), sugar alcohols (e.g., glycerol), ascorbic acid intermediates (e.g., gluconate, 2-keto-D-gluconate, 2,5-diketo-D-gluconate, and 2-keto-L-gulonic acid), amino acids (e.g., lysine), proteins (e.g., antibodies and fragment thereof).

Paragraph 48. An isolated polynucleotide encoding the vari- 15 ant of any of paragraphs 1-33.

Paragraph 49. A nucleic acid construct comprising the polynucleotide of paragraph 48.

Paragraph 50. An expression vector comprising the nucleic acid construct of paragraph 49.

Paragraph 51. A host cell comprising the nucleic acid construct of paragraph 49.

Paragraph 52. A method of producing a variant, comprising: a. cultivating the host cell of paragraph 51 under conditions suitable for the expression of the alpha-amylase; and

b. recovering the variant from the cultivation medium. Paragraph 53. A transgenic plant, plant part or plant cell transformed with the polynucleotide of paragraph 48. Paragraph 54. A method for preparing a variant of a parent alpha-amylase comprising the following steps:

a. providing a nucleic acid encoding a parent alpha-amy-lase.

b. introducing alterations in the nucleic acid sequence resulting in alterations of the encoded amino acid residues in two, three, four or five positions, said positions corresponding 35 to positions in the parent alpha-amylase selected from the group consisting of 163, 188, 205, 208 and 209; the alteration(s) are independently

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- (i) an insertion of an amino acid immediately downstream of the position,
- (ii) a deletion of the amino acid which occupies the position, and/or
- (iii) a substitution of the amino acid which occupies the position,
- c. optionally introducing further alterations in the nucleic acid sequence resulting in alterations of one or more amino acid residues in the B-domain of the parent alpha-amylase to the corresponding amino acid residue(s) in SEQ ID NO: 9;

d. optionally introducing further alterations in the nucleic acid sequence resulting in alterations of the encoded amino acid residues selected from the group of X183\*+X184\*, X186A,Y,T, X193F, X195F, M202L,I,T,S,A, X206F,Y, X214I, X244A,D,E,N,Q,W, X452H,Y, X474R and X475R;

e. expressing the altered nucleic acid sequence in a suitable host organism; and

f. recovering the variant alpha-amylase,

wherein each position corresponds to a position of the amino acid sequence of the enzyme having the amino acid sequence of SEQ ID NO: 2.

Paragraph 55. The method of paragraph 54, wherein the alterations in b. are selected among: X163Q,N, X188N, X205N, X208Y and, X209N,S.

Paragraph 56. The method of paragraph 54 or 55, wherein the parent alpha-amylase is a Termamyl-like alpha-amylase. Paragraph 57. The method of paragraph 56, wherein the parent alpha-amylase is selected among alpha-amylases having SEQ ID NO: 2, 4, 6, 8 or 10, or alpha-amylases derived from a strain of the *Bacillus* sp. NCIB 12289, NCIB 12512, NCIB 12513 or DSM 9375, and the #707 alpha-amylase described by Tsukamoto et al., 1988, *Biochemical and Biophysical Research Communications* 151: 25-31, or alpha-amylases having at least 70%, preferably at least 75%, more preferably at least 80%, more preferably at least 95%, and even most preferably at least about 97% sequence identity to the amino acid sequence of one of these.

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Thr Arg Glu Ser Gly Tyr Pro Gln Val Phe Tyr Gly Asp Met Tyr Gly 350  Thr Lys Gly Asp Ser Gln Arg Glu Ile Pro Ala Leu Lys His Lys Ile 370  Glu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His 385  Glu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His 385  Ser Ser Val Ala Asn Ser Gly Leu Ala Ala Leu Ile Thr Asp Glu Gly Asp 405  Ser Ser Val Ala Asn Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro 420  Gly Gly Ala Lys Arg Met Tyr Val Gly Arg Gln Asn Ala Gly Glu Thr 445  Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser 450  Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 470  Val Gln Arg  **210> SEO ID NO 5  **2211> LENGTH: 1452  **2212> TYPE: DNA  **2122> TYPE: DNA  **213> ORGANISM: Bacillus amyloliquefaciens  **222> DeATION: (1)(1452)  **400> SEQUENCE: 5  gda aat ggc acg ctg atg cag tat ttt gaa tgg tat acg ccg aac gac Val Asn Gly Gln His Trp Lys Arg Leu Gln Asn Asp Ala Glu His Leu Ser Asp 20  atc gga atc act ggc acg ttg ag aat gat gat gg gaa cat tta tcg gat Gly Gln His Trp Lys Arg Leu Gln Asn Asp Ala Glu His Leu Ser Asp 20  atc gga atc act gcc gtc tg atg cat cct ccc gca tac aaa gga ttg agc 144  Ile Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Tyr Lys Gly Leu Ser 35  caa tcc gat aac gga tac gga cct tat gat ttg tat ta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat ta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat ggt tta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat ta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat gat tta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat ta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat ta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat gat tta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat gat tta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat gat tta gga gaa [192]  caa tcc gat aac gga tac gga cct tat gat ttg tat gat tta gga [192]  cat cag caa aaa gga acg gtc aga aca aaa tcc gga aca aaa tca gag [24	Val	Thr	Phe	Val							Pro	Gly	Gln	Ser		Glu		
### The Lys Gly Asp Ser Gln Arg Glu Ile Pro Ala Leu Lys His Lys Ile 370	Ser	Thr	Val		Thr	Trp	Phe	Lys		Leu					Ile	Leu		
Glu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His 385	Thr	Arg							Val						Tyr	Gly		
Clu Pro Ile Leu Lys Ala Arg Lys Gln Tyr Ala Tyr Gly Ala Gln His 385   390   390   395   400   Asp 405   410   415   415	Thr			Asp	Ser	Gln	Arg	Glu	Ile	Pro	Ala		ГÀа	His	Lys	Ile		
Asp Tyr Phe Asp His His Asp Ile Val Gly Trp Thr Arg Glu Gly Asp 405  Ser Ser Val Ala Asn Ser Gly Leu Ala Ala Leu Ile Thr Asp Gly Pro 420  Gly Gly Ala Lys Arg Met Tyr Val Gly Arg Gln Asn Ala Gly Glu Thr 435  Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser 450  Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 465  Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 465  470  Val Gln Arg <pre> &lt;210&gt; SEQ ID NO 5</pre>		Pro				Ala	Arg	Lys	Gln	Tyr		Tyr	Gly	Ala	Gln			
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Gly Gly Ala Lys Arg Met Tyr Val Gly Arg Gln Asn Ala Gly Glu Thr  435  Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser  450  Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 465  470  Val Gln Arg <pre> <a href="#"> <a hre="#"> <a href="#"> <a href="#"> <a href="#"> <a href="#"> <a href<="" td=""><td>Ser</td><td>Ser</td><td>Val</td><td></td><td></td><td>Ser</td><td>Gly</td><td>Leu</td><td></td><td></td><td></td><td>Ile</td><td>Thr</td><td></td><td></td><td>Pro</td><td></td><td></td></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></pre>	Ser	Ser	Val			Ser	Gly	Leu				Ile	Thr			Pro		
Trp His Asp Ile Thr Gly Asn Arg Ser Glu Pro Val Val Ile Asn Ser 450  Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 465 470  Val Gln Arg <pre> <a href="#"> <a href="#"></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></pre>	Gly	Gly			Arg	Met	Tyr			Arg	Gln	Asn			Glu	Thr		
Glu Gly Trp Gly Glu Phe His Val Asn Gly Gly Ser Val Ser Ile Tyr 465 470 475 480  Val Gln Arg <pre> <a href="#"> </a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></a></pre>																		

						51					0.0	- ,-	,,,,	_,,		52	
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				85					90					95			
						cat His										336	
-		_	_	_	_	aat Asn	_	_		_		_	_		_	384	
						gcg Ala 135										432	
						ttt Phe										480	
						cgg Arg										528	
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						gct Ala										624	
						tgg Trp 215										672	
						gat Asp										720	
						gcg Ala										768	
						tgg Trp										816	
						ttt Phe										864	
						gct Ala 295										912	
	_	_	_	_	-	acc Thr	_					_	_	_		960	
						cat His										1008	
_		_				ttt Phe		_		_		_			_	1056	
	-	-				cct Pro	_					-	_			1104	
						aag Lys 375										1152	
						cgt Arg										1200	

gat tat att gac cac ccg gat gtg atc gga tgg acg agg gaa ggt gac

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tgg tat gac ata acg gg Trp Tyr Asp Ile Thr Gl 450	y Asn Arg Ser Asp Thr		1392
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Ile Gly Ile Thr Ala Va 35	l Trp Ile Pro Pro Ala 40	Tyr Lys Gly Leu Ser 45	
Gln Ser Asp Asn Gly Ty 50		Tyr Asp Leu Gly Glu 60	
Phe Gln Gln Lys Gly Th 65 70		Gly Thr Lys Ser Glu 80	
Leu Gln Asp Ala Ile Gl 85	y Ser Leu His Ser Arg 90	Asn Val Gln Val Tyr 95	
Gly Asp Val Val Leu As 100	n His Lys Ala Gly Ala 105	Asp Ala Thr Glu Asp 110	
Val Thr Ala Val Glu Va 115	l Asn Pro Ala Asn Arg 120	Asn Gln Glu Thr Ser 125	
Glu Glu Tyr Gln Ile Ly 130		Arg Phe Pro Gly Arg 140	
Gly Asn Thr Tyr Ser As 145 15		Tyr His Phe Asp Gly 160	
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Gly Glu Gly Lys Ala Tr 180	p Asp Trp Glu Val Ser 185	Ser Glu Asn Gly Asn 190	
Tyr Asp Tyr Leu Met Ty 195	r Ala Asp Val Asp Tyr 200	Asp His Pro Asp Val 205	
Val Ala Glu Thr Lys Ly 210		Ala Asn Glu Leu Ser 220	
Leu Asp Gly Phe Arg Il 225 23		Ile Lys Phe Ser Phe 240	
Leu Arg Asp Trp Val Gl 245	n Ala Val Arg Gln Ala 250	Thr Gly Lys Glu Met 255	
Phe Thr Val Ala Glu Ty	r Trp Gln Asn Asn Ala	Gly Lys Leu Glu Asn	

											COII	CIII	uea		
		260					265					270			
Tyr Leu	1 Asn 275	-	Thr	Ser	Phe	Asn 280	Gln	Ser	Val	Phe	Asp 285	Val	Pro	Leu	
His Phe		Leu	Gln	Ala	Ala 295	Ser	Ser	Gln	Gly	Gly 300	Gly	Tyr	Asp	Met	
Arg Arg 305	g Leu	. Leu	Asp	Gly 310		Val	Val	Ser	Arg 315		Pro	Glu	Lys	Ala 320	
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Thr Arg	g Glu 355		Gly	Tyr	Pro	Gln 360	Val	Phe	Tyr	Gly	Asp 365	Met	Tyr	Gly	
Thr Lys		Thr	Ser	Pro	Lys 375	Glu	Ile	Pro	Ser	Leu 380	Lys	Asp	Asn	Ile	
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Asp Tyı	: Ile	Asp	His 405	Pro	Asp	Val	Ile	Gly 410	Trp	Thr	Arg	Glu	Gly 415	Asp	
Ser Sei	Ala	Ala 420		Ser	Gly	Leu	Ala 425	Ala	Leu	Ile	Thr	Asp 430	Gly	Pro	
Gly Gl	/ Ser 435	_	Arg	Met	Tyr	Ala 440	Gly	Leu	Lys	Asn	Ala 445	Gly	Glu	Thr	
Trp Tyr	_	Ile	Thr	Gly	Asn 455	Arg	Ser	Asp	Thr	Val 460	Lys	Ile	Gly	Ser	
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Val Glr	ı Lys														
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1			5	•				10	•			-	15		
ccg gat Pro Asp															96
tta tco Leu Sei															144
gga aca Gly Thi															192
ctc ggd Leu Gl <sub>y</sub> 65															240
aaa gct Lys Ala															288
caa gtg	g tac	gcc	gat	gtc	gtg	ttc	gac	cat	aaa	ggc	ggc	gct	gac	ggc	336

Gin Val Tyr Ala Asp Val Val Phe Asp His Lye Gly Gly Ala Asp Gly 100 100 100 105 100 105 100 105 100 105 100 105 100 105 100 105 100 105 125 125 125 125 125 125 125 125 125 12																	
The fall Tip Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln 115 125 120 125 125 125 125 125 125 125 125 125 125	Gln	Val	Tyr		Asp	Val	Val	Phe	_	His	Lys	Gly	Gly		Asp	Gly	
Glu Ile Ser Gly Thr Tyr Gln 1le Gln Ala Trp Thr Lyp Phe Asp Phe 130 135 136 136 146 156 156 166 130 166 130 166 130 166 130 166 130 166 130 166 130 166 155 166 166 166 166 166 166 166 166			Trp					Glu					Asp				384
Pro Gily Arg Gily Asm Thr Tyr Ser Ser Phe Lye Trp Arg Trp Tyr His 160		Ile					Gln					Thr					432
Phe Asp Giy Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr 175  asa ttc cgc ggc atc ggc aaa gcg tgg gat tgg gaa gta gac acg gaa Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu 180  acac gga aac tat gac tac tta atg tat gcc gac ctt gat atg gat cat Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His 200  ccc gaa gtc gtg acc gag ctg aaa aac tgg ggg aaa tgg tat gtc aac Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn 210  aca acg aac att gat ggg ttc cgg ctt gat ggc gtc aag cat at aag Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys 230  ctc agt ttt ttt cct gat tgg ttg cg tat ggc gtc aag cat att aag Phe Ser Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly 245  aag ccg cta ttt acc gtc ggg gaa tat tgg agc tat gac act ggc Phe Ser Phe Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys 266  ttg cac aat tac att acg aaa aca gac ggg acg at ttg gac atc aac aag Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys 275  gcc ccg tta cac aac aaa ttt tat acc gct tcc aaa tca ggg gg gc a La Lys Asn Tyr Ile Thr Lys Thr Asp Gly Thr Met Ser Leu Phe Asp 275  gcc ccg tta cac aac aaa ttt tat acc gct tcc aaa tca ggg gg gc a Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala 295  ctt gat atg cgc acg tta atg acc aat act ctc atg asa gat cac ge Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro 305  aca ttg gcc gtc acc ttc gtt gat aat cat gac acc gac ccg caa 1008  Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln 330  gcg ctg cag tca tcg acg ga gga tat ccg ttc aac acc gg gcc caa 1104  Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 345  ctt att cta act cgg cag ga gga tat ccg ttc aaa acc gt ggc cac acc acc ggc cac acc acc acc acc	Pro					Thr					Lys					His	480
Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu 180  aac gga aac tat gac tac tta atg tat gcc gac ctt gat atg gat cat 624 Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His 200  ccc gaa gtc gtg acc gag ctg aaa aac tgg ggg aaa tgg tat gtc aac pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn 210  aca acg aac att gat ggg ttc cgg ctt gat gcc gtc aag cat att aag 720  tro Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys 230  ttc agt ttt ttc cct gat tgg ttg tcg tat gtg cgt tct cag act ggc Phe Ser Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly 255  aag ccg cta ttt acc gtc ggg gaa tat tgg acc tat gac atc aac aag Lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys 260  ttg cac aat tac att acc gac acc gac gga acg atg tct ttg He Asp Ala Pro Leu His Asn Lys Phe Tyr Thr Asp Gly Thr Met Ser Leu Phe Asp 275  gcc ccg tta cac aac aaa ttt tat acc gct tcc aat acc agg ggc gcc ala Phe Asp Net Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro 310  ttt gat atg cgc acg tta atg acc aat act ctc atg aaa aga cac ccg gc caa 1008  ttt gat atg cgc acg tta atg acc aat act ctc atg aaa aga cac ccg gc caa 1008  Thr Leu Ala Val Thr Phe Val Asp Asp His Asp Thr Glu Pro Gly Gln 310  aca ttg gcc gtc acc ttc gtt gat aat cat gac acc gaa ccc ggc caa 1008  Thr Leu Ala Val Thr Phe Val Asp Asp His Asp Thr Glu Pro Gly Gln 320  aca ttg gcc gtc acc ttc gtt gat aat cat gac acc gaa ccc ggc caa 1008  Thr Leu Ala Val Thr Phe Val Asp Asp His Asp Thr Glu Pro Gly Gln 320  aca ttg gcc gtc acc ttc gtt gat acc cat ggt tc aac cg gt gc cac 1104  Thr Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Ala 320  aca ttg gcc dcd cac aca at at acc acc tgg ttc aca acc gac ccc ggc caa 1008  Thr Leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp 355  acc gcc cc ctc ctc atc gcc gc agg gat tat gct tac gga acc cac acc acc acc acc acc acc a					Asp					Arg					Ile		528
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Pro			Asn					Met					Asp				624
Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys 235  ttc agt ttt ttt cot gat tgg ttg tog tag gtg gt cot cag act ggc Phe Ser Phe Phe Pro Asp Trp Leu Ser Tyr Val Arg Ser Gln Thr Gly 245  aag cog cta ttt acc gtc ggg gaa tat tgg agc tat gac atc aac aag lys Pro Leu Phe Thr Val Gly Glu Tyr Trp Ser Tyr Asp Ile Asn Lys 260  ttg cac aat tac att acg aaa aca gac gga acg atg ttt ttg ttt gat Leu His Asn Tyr Ile Thr Lys Thr Asp Gly Thr Met Ser Leu Phe Asp 275  gcc ccg tta cac aac aaa ttt tat acc gct tc aaa tca ggg ggc gca Ala Pro Leu His Asn Lys Phe Tyr Thr Ala Ser Lys Ser Gly Gly Ala 290  ttt gat atg cgc acg tta atg acc aat act ct aga at caa ccg Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro 310  aca ttg gcc gtc acc ttc gtt gat aat cat gac acc gac gac acc gac cgc cad Thr Leu Ala Val Thr Phe Val Asp Ash His Asp Thr Glu Pro Gly Gln Asp 330  gcg ctg cag tca tgg gtc gac cca tgg ttc aaa ccg ttg gct tac gcc Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 325  gcg ctg cag tca ccg tca gas cca tgg ttc aaa ccg ttg gct tac gcc Ala Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 340  ttt att cta act cgg cag gaa tac ccg tgg gtc ttt tat ggt gac losd Leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 355  ttt att ggc atc cac cac at acc acc acc tgg ttc aaa ccg ttg gct tac gcc losd acc gcc cac gcc cac gas acc gac cac gcc cac ca		Glu					Leu					Lys					672
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Second   S				Phe					Tyr					Ile			816
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Phe Asp Met Arg Thr Leu Met Thr Asn Thr Leu Met Lys Asp Gln Pro 320  aca ttg gcc gtc acc ttc gtt gat aat cat gac acc gaa ccc ggc caa Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln 335  gcg ctg cag tca tgg gtc gac cca tgg ttc aaa ccg ttg gct tac gcc loss and leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 340  ttt att cta act cgg cag gaa gga tac ccg tgc gtc ttt tat ggt gac loss and leu Thr Arg Gln Glu Gly Tyr Pro Cys Val Phe Tyr Gly Asp 355  tat tat ggc att cca caa tat aac att cct tcg ctg aaa agc aaa atc loss and leu Lys Ser Lys Ile 370  gat ccg ctc ctc atc gcg cag agg gat tat gct tac gga acg caa cat Asp Pro Leu Leu Lys Ser Lys Ile 385  gat ccg ctc ctc atc gcg cag agg gat tat gct tac gga acg caa cat loss app Pro Leu Leu Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His 395  gat tat ctt gat cac tcc gac atc atc ggg tgg tgg aca agg gaa ggg ggc loss app Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Gly	_	Pro					Phe			_		Lys				_	912
Thr Leu Ala Val Thr Phe Val Asp Asn His Asp Thr Glu Pro Gly Gln 325    gcg ctg cag tca tgg gtc gac cca tgg ttc aaa ccg ttg gct tac gcc loss and leu Gln Ser Trp Val Asp Pro Trp Phe Lys Pro Leu Ala Tyr Ala 340    ttt att cta act cgg cag gaa gga tac ccg tgc gtc ttt tat ggt gac loss as 350    ttt att cta act cgg cag gaa gga tac ccg tgc gtc ttt tat ggt gac loss as 350    tat tat ggc att cca caa tat aac att cct tcg ctg aaa agc aaa atc loss as 360    tat tat ggc att cca caa tat aac att cct tcg ctg aaa agc aaa atc loss as 375    gat ccg ctc ctc atc gcg cgc agg gat tat gct tac gga acg caa cat Asp Pro Leu Leu Lus Ser Lys Ile 370    gat ccg ctc ctc atc gcg cgc agg gat tat gct tac gga acg caa cat Asp Pro Leu Leu Lus Ile Ala Arg Arg Asp Tyr Ala Tyr Gly Thr Gln His 385    gat tat ctt gat cac tcc gac atc atc ggg tgg aca agg gaa ggg ggc loss as 395    gat tat ctt gat cac tcc gac atc atc ggg tgg aca agg gaa ggg ggc loss as 395    gat tat ctt gat cac tcc gac atc atc ggg tgg aca agg gaa ggg ggc loss as 395    gat tat ctt gat cac tcc gac atc atc ggg tgg aca agg gaa ggg ggc loss as 395    loss acc ggg ggc loss agg ggc loss agg ggg loss agg gga ggg ggc loss agg Tyr Leu Asp His Ser Asp Ile Ile Gly Trp Thr Arg Glu Gly Gly	Phe	_	_	_	_	Leu	_				Leu	_		_		Pro	960
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Ala	Thr	Glu 115	Asn	Val	Leu	Ala	Val 120	Glu	Val	Asn	Pro	Asn 125	Asn	Arg	Asn
Gln	Glu 130	Ile	Ser	Gly	Asp	Tyr 135	Thr	Ile	Glu	Ala	Trp 140	Thr	Lys	Phe	Asp
Phe 145	Pro	Gly	Arg	Gly	Asn 150	Thr	Tyr	Ser	Asp	Phe 155	ГÀа	Trp	Arg	Trp	Tyr 160
His	Phe	Asp	Gly	Val 165	Asp	Trp	Asp	Gln	Ser 170	Arg	Gln	Phe	Gln	Asn 175	Arg
Ile	Tyr	Lys	Phe 180	Arg	Gly	Asp	Gly	Lys 185	Ala	Trp	Asp	Trp	Glu 190	Val	Asp
Ser	Glu	Asn 195	Gly	Asn	Tyr	Asp	Tyr 200	Leu	Met	Tyr	Ala	Asp 205	Val	Asp	Met
Asp	His 210	Pro	Glu	Val	Val	Asn 215	Glu	Leu	Arg	Arg	Trp 220	Gly	Glu	Trp	Tyr
Thr 225	Asn	Thr	Leu	Asn	Leu 230	Asp	Gly	Phe	Arg	Ile 235	Asp	Ala	Val	Lys	His 240
Ile	Lys	Tyr	Ser		Thr			Trp			His	Val	Arg	Asn 255	
Thr	Gly	Lys	Glu 260	Met	Phe	Ala	Val	Ala 265	Glu	Phe	Trp	Lys	Asn 270	Asp	Leu
Gly	Ala	Leu 275	Glu	Asn	Tyr	Leu	Asn 280	ГЛа	Thr	Asn	Trp	Asn 285	His	Ser	Val
Phe	Asp 290	Val	Pro	Leu	His	Tyr 295	Asn	Leu	Tyr	Asn	Ala 300	Ser	Asn	Ser	Gly
Gly 305	Asn	Tyr	Asp	Met	Ala 310	Lys	Leu	Leu	Asn	Gly 315	Thr	Val	Val	Gln	Lys 320
His	Pro	Met	His	Ala 325	Val	Thr	Phe	Val	Asp 330	Asn	His	Asp	Ser	Gln 335	Pro
Gly	Glu	Ser	Leu 340	Glu	Ser	Phe	Val	Gln 345	Glu	Trp	Phe	Lys	Pro 350	Leu	Ala
Tyr	Ala	Leu 355	Ile	Leu	Thr	Arg	Glu 360	Gln	Gly	Tyr	Pro	Ser 365	Val	Phe	Tyr

Gly Asp Tyr Tyr Gly Ile Pro Thr His Ser Val Pro Ala Met Lys Ala 370 375 380

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Lys Ile Asp Pro Ile Leu Glu Ala Arg Gln Asn Phe Ala Tyr Gly Thr 390 Gln His Asp Tyr Phe Asp His His Asn Ile Ile Gly Trp Thr Arg Glu Gly Asn Thr Thr His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp 425 Gly Pro Gly Gly Glu Lys Trp Met Tyr Val Gly Gln Asn Lys Ala Gly Gln Val Trp His Asp Ile Thr Gly Asn Lys Pro Gly Thr Val Thr Ile Asn Ala Asp Gly Trp Ala Asn Phe Ser Val Asn Gly Gly Ser Val Ser Ile Trp Val Lys Arg <210> SEQ ID NO 11 <211> LENGTH: 480 <212> TYPE: PRT <213 > ORGANISM: Bacillus sp. <400> SEQUENCE: 11 Asp Gly Leu Asn Gly Thr Met Met Gln Tyr Tyr Glu Trp His Leu Glu Asn Asp Gly Gln His Trp Asn Arg Leu His Asp Asp Ala Ala Ala Leu Ser Asp Ala Gly Ile Thr Ala Ile Trp Ile Pro Pro Ala Tyr Lys Gly 40 Asn Ser Gln Ala Asp Val Gly Tyr Gly Ala Tyr Asp Leu Tyr Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys Ala Gln Leu Glu Arg Ala Ile Gly Ser Leu Lys Ser Asn Asp Ile Asn Val Tyr Gly Asp Val Val Met Asn His Lys Met Gly Ala Asp Phe Thr Glu Ala Val Gln Ala Val Gln Val Asn Pro Thr Asn Arg Trp Gln Asp Ile Ser Gly Ala Tyr Thr Ile Asp Ala Trp Thr Gly Phe Asp Phe Ser Gly Arg Asn Asn Ala Tyr Ser Asp Phe Lys Trp Arg Trp Phe His Phe Asn Gly Val Asp Trp Asp Gln Arg Tyr Gln Glu Asn His Ile Phe Arg Phe Ala Asn Thr Asn Trp Asn Trp Arg Val Asp Glu Glu Asn Gly Asn Tyr Asp Tyr Leu Leu Gly Ser Asn Ile Asp Phe Ser His Pro Glu Val 200 Gln Asp Glu Leu Lys Asp Trp Gly Ser Trp Phe Thr Asp Glu Leu Asp 215 Leu Asp Gly Tyr Arg Leu Asp Ala Ile Lys His Ile Pro Phe Trp Tyr Thr Ser Asp Trp Val Arg His Gln Arg Asn Glu Ala Asp Gln Asp Leu 250 Phe Val Val Gly Glu Tyr Trp Lys Asp Asp Val Gly Ala Leu Glu Phe 265

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Tyr Leu Asp Glu Met Asn Trp Glu Met Ser Leu Phe Asp Val Pro Leu 280 Asn Tyr Asn Phe Tyr Arg Ala Ser Gln Gln Gly Gly Ser Tyr Asp Met 295 Arg Asn Ile Leu Arg Gly Ser Leu Val Glu Ala His Pro Met His Ala Val Thr Phe Val Asp Asn His Asp Thr Gln Pro Gly Glu Ser Leu Glu Ser Trp Val Ala Asp Trp Phe Lys Pro Leu Ala Tyr Ala Thr Ile Leu Thr Arg Glu Gly Gly Tyr Pro Asn Val Phe Tyr Gly Asp Tyr Tyr Gly 355 360 365 Ile Pro Asn Asp Asn Ile Ser Ala Lys Lys Asp Met Ile Asp Glu Leu Leu Asp Ala Arg Gln Asn Tyr Ala Tyr Gly Thr Gln His Asp Tyr Phe 395 Asp His Trp Asp Val Val Gly Trp Thr Arg Glu Gly Ser Ser Ser Arg 405  $\phantom{\bigg|}405$ Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asn Gly Pro Gly Gly Ser 425 Lys Trp Met Tyr Val Gly Arg Gln Asn Ala Gly Gln Thr Trp Thr Asp 440 Leu Thr Gly Asn Asn Gly Ala Ser Val Thr Ile Asn Gly Asp Gly Trp 455 Gly Glu Phe Phe Thr Asn Gly Gly Ser Val Ser Val Tyr Val Asn Gln 470 475 <210> SEQ ID NO 12 <211> LENGTH: 586 <212> TYPE: PRT <213 > ORGANISM: Bacillus <400> SEQUENCE: 12 Gly Ser Val Pro Val Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Asn Ala Gln 25 Ser Leu Ala Asn Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys Gly Thr Ser Ser Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys Thr Gln Tyr Ile Gln Ala Ile Gln Ala Ala His Thr Ala Gly Met Gln Val Tyr Ala Asp Val Val Phe Asn His Lys Ala Gly Ala Asp 105 Gly Thr Glu Leu Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn 120 Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His Phe Asp Gly Thr Asp Trp Asp Glu Ser Arg Lys Leu Asn Arg Ile

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													C III.		
				165					170					175	
Tyr L	Хa	Phe	Arg 180	Gly	Thr	Gly	Lys	Ala 185	Trp	Asp	Trp	Glu	Val 190	Asp	Thr
Glu A	sn	Gly 195	Asn	Tyr	Asp	Tyr	Leu 200	Met	Tyr	Ala	Asp	Leu 205	Asp	Met	Asp
His P	ro 10	Glu	Val	Val	Ser	Glu 215	Leu	Lys	Asn	Trp	Gly 220	Lys	Trp	Tyr	Val
Ile T 225	hr	Thr	Asn	Ile	Asp 230	Gly	Phe	Arg	Leu	Asp 235	Ala	Val	ГÀз	His	Ile 240
Lys T	yr	Ser	Phe	Phe 245	Pro	Asp	Trp	Leu	Ser 250	Tyr	Leu	Arg	Thr	Gln 255	Thr
Gln L	Уs	Pro	Leu 260	Phe	Ala	Val	Gly	Glu 265	Phe	Trp	Ser	Tyr	Asp 270	Ile	Asn
Lys L	eu	His 275	Asn	Tyr	Ile	Thr	Lys 280	Thr	Asn	Gly	Ser	Met 285	Ser	Leu	Phe
Asp A	la 90	Pro	Leu	His	Asn	Asn 295	Phe	Tyr	Ile	Ala	Ser 300	Lys	Ser	Gly	Gly
Tyr P 305	he	Asp	Met	Arg	Thr 310	Leu	Leu	Asn	Asn	Thr 315	Leu	Met	Lys	Glu	Gln 320
Pro T	hr	Leu	Ser	Val 325	Thr	Leu	Val	Asp	Asn 330	His	Asp	Thr	Glu	Pro 335	Gly
Gln S	er	Leu	Gln 340	Ser	Trp	Val	Glu	Pro 345	Trp	Phe	Lys	Pro	Leu 350	Ala	Tyr
Ala P	he	Ile 355	Leu	Thr	Arg	Gln	Glu 360	Gly	Tyr	Pro	Cys	Val 365	Phe	Tyr	Gly
Asp T	yr 70	Tyr	Gly	Ile	Pro	Lys 375	Tyr	Asn	Ile	Pro	Ala 380	Leu	Lys	Ser	Lys
Leu A 385	ap	Pro	Leu	Leu	Ile 390	Ala	Arg	Arg	Asp	Tyr 395	Ala	Tyr	Gly	Thr	Gln 400
His A	ap	Tyr	Ile	Asp 405	Asn	Ala	Asp	Ile	Ile 410	Gly	Trp	Thr	Arg	Glu 415	Gly
Val A	la	Glu	Lys 420	Ala	Asn	Ser	Gly	Leu 425	Ala	Ala	Leu	Ile	Thr 430	Asp	Gly
Pro G	ly	Gly 435	Ser	Lys	Trp	Met	Tyr 440	Val	Gly	Lys	Gln	His 445	Ala	Gly	Lys
Thr P	he 50	Tyr	Asp	Leu	Thr	Gly 455		Arg	Ser	Asp	Thr 460		Thr	Ile	Asn
Ala A 465	ap	Gly	Trp	Gly	Glu 470	Phe	Lys	Val	Asn	Gly 475	Gly	Ser	Val	Ser	Ile 480
Trp V	al	Pro	Lys	Thr 485	Ser	Thr	Thr	Ser	Gln 490	Ile	Thr	Phe	Thr	Val 495	Asn
Asn A	la	Thr	Thr 500	Val	Trp	Gly	Gln	Asn 505	Val	Tyr	Val	Val	Gly 510	Asn	Ile
Ser G	ln	Leu 515	Gly	Asn	Trp	Asp	Pro 520	Val	Asn	Ala	Val	Gln 525	Met	Thr	Pro
Ser S 5	er 30	Tyr	Pro	Thr	Trp	Val 535	Val	Thr	Val	Pro	Leu 540	Pro	Gln	Ser	Gln
Asn I 545	le	Gln	Phe	Lys	Phe 550	Ile	Lys	Lys	Asp	Gly 555	Ser	Gly	Asn	Val	Ile 560
Trp G	lu	Asn	Ile	Ser 565	Asn	Arg	Thr	Tyr	Thr 570	Val	Pro	Thr	Ala	Ala 575	Ser
Gly A	la	Tyr	Thr 580	Ala	Asn	Trp	Asn	Val 585	Pro						

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Ile Asp Pro Le	an I.an	Tla	7. T. a	Ara	Ara	Δen	Тчи	7. T a	Туг	G1 v	Thr	Gln
385	ва пеа	390	AIA	AIG	AIG	Yeb	395	AIA	ıyı	GIY	1111	400
Arg Asp Tyr I	le Asp 405	His	Gln	Asp	Ile	Ile 410	Gly	Trp	Thr	Arg	Glu 415	Gly
Ile Asp Thr Ly	ys Pro 20	Asn	Ser	Gly	Leu 425	Ala	Ala	Leu	Ile	Thr 430	Asp	Gly
Pro Gly Gly Se 435	er Lys	Trp	Met	Tyr 440	Val	Gly	Lys	Lys	His 445	Ala	Gly	Lys
Val Phe Tyr As 450	sp Leu	Thr	Gly 455	Asn	Arg	Ser	Asp	Thr 460	Val	Thr	Ile	Asn
Ala Asp Gly Tr 465	rp Gly	Glu 470	Phe	Lys	Val	Asn	Gly 475	Gly	Ser	Val	Ser	Ile 480
Trp Val Ala Ly	ys Thr 485	Ser	Asn	Val	Thr	Phe 490	Thr	Val	Asn	Asn	Ala 495	Thr
Thr Thr Ser G	ly Gln 00	Asn	Val	Tyr	Val 505	Val	Ala	Asn	Ile	Pro 510	Glu	Leu
Gly Asn Trp As 515	sn Thr	Ala	Asn	Ala 520	Ile	Lys	Met	Asn	Pro 525	Ser	Ser	Tyr
Pro Thr Trp Ly 530	ys Ala	Thr	Ile 535	Ala	Leu	Pro	Gln	Gly 540	Lys	Ala	Ile	Glu
Phe Lys Phe I 545	le Lys	550 Lys	Asp	Gln	Ala	Gly	Asn 555	Val	Ile	Trp	Glu	Ser 560
Thr Ser Asn A	rg Thr 565	Tyr	Thr	Val	Pro	Phe 570	Ser	Ser	Thr	Gly	Ser 575	Tyr
Thr Ala Ser Ti	rp Asn 30	Val	Pro									
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	485 RT	illus	5									
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<pre>&lt;211&gt; LENGTH: &lt;212&gt; TYPE: PI &lt;213&gt; ORGANIST &lt;400&gt; SEQUENCE His His Asn G 1 Leu Pro Asn As 20 Asn Leu Lys S 35 Lys Gly Thr S 50 Asp Leu Gly G: 65 Thr Arg Ser G: Ile Gln Val T</pre>	485 RT W: Bac E: 14 Ly Thr S SP Gly D ET Lys Lu Phe Lu Phe Lu R5 Vr Gly D0	Asn Gly Asn Asn Gln Asp	Gly His Ile Asp 55 Gln Gly Val	Trp Thr 40 Val Lys Ala	Asn 25 Ala Gly Val Met 105	10 Arg Val Tyr Thr Asn	Leu Trp Gly Val 75 Ser	Arg Ile Ala 60 Arg Leu Lys	Asp Pro 45 Tyr Thr Lys	Asp 30 Pro Asp Lys Asn Gly 110	Ala Ala Leu Tyr Asn 95 Ala	Ala Trp Tyr Gly 80 Gly Asp
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His Phe Asp Gly Thr Asp Trp Asp Gln Ser Arg Gln Leu Gln Asn Lys 165 170 Ile Tyr Lys Phe Arg Gly Thr Gly Lys Ala Trp Asp Trp Glu Val Asp Ile Glu Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Met Asp His Pro Glu Val Ile Asn Glu Leu Arg Asn Trp Gly Val Trp Tyr Thr Asn Thr Leu Asn Leu Asp Gly Phe Arg Ile Asp Ala Val Lys His Ile Lys Tyr Ser Tyr Thr Arg Asp Trp Leu Thr His Val Arg Asn Thr Thr Gly Lys Pro Met Phe Ala Val Ala Glu Phe Trp Lys Asn Asp Leu Ala Ala Ile Glu Asn Tyr Leu Asn Lys Thr Ser Trp Asn His Ser Val Phe Asp Val Pro Leu His Tyr Asn Leu Tyr Asn Ala Ser Asn Ser Gly 290 295 300 Gly Tyr Phe Asp Met Arg Asn Ile Leu Asn Gly Ser Val Val Gln Lys 310 His Pro Ile His Ala Val Thr Phe Val Asp Asn His Asp Ser Gln Pro 325 330 Gly Glu Ala Leu Glu Ser Phe Val Gln Ser Trp Phe Lys Pro Leu Ala 345 Tyr Ala Leu Ile Leu Thr Arg Glu Gln Gly Tyr Pro Ser Val Phe Tyr Gly Asp Tyr Tyr Gly Ile Pro Thr His Gly Val Pro Ser Met Lys Ser 375 Lys Ile Asp Pro Leu Leu Gln Ala Arg Gln Thr Tyr Ala Tyr Gly Thr 390 Gln His Asp Tyr Phe Asp His His Asp Ile Ile Gly Trp Thr Arg Glu Gly Asp Ser Ser His Pro Asn Ser Gly Leu Ala Thr Ile Met Ser Asp Gly Pro Gly Gly Asn Lys Trp Met Tyr Val Gly Lys His Lys Ala Gly 440 Gln Val Trp Arg Asp Ile Thr Gly Asn Arg Ser Gly Thr Val Thr Ile Asn Ala Asp Gly Trp Gly Asn Phe Thr Val Asn Gly Gly Ala Val Ser 465  $\phantom{\bigg|}470\phantom{\bigg|}470\phantom{\bigg|}475\phantom{\bigg|}475\phantom{\bigg|}$ Val Trp Val Lys Gln <210> SEQ ID NO 15 <211> LENGTH: 485 <212> TYPE: PRT <213 > ORGANISM: Bacillus <400> SEQUENCE: 15 His His Asn Gly Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu Pro Asn Asp Gly Asn His Trp Asn Arg Leu Arg Ser Asp Ala Ser 25 Asn Leu Lys Asp Lys Gly Ile Thr Ala Val Trp Ile Pro Pro Ala Trp

Lys	Gly 50	Ala	Ser	Gln	Asn	Asp 55	Val	Gly	Tyr	Gly	Ala 60	Tyr	Asp	Leu	Tyr
Asp 65	Leu	Gly	Glu	Phe	Asn 70	Gln	Lys	Gly	Thr	Val 75	Arg	Thr	Lys	Tyr	Gly 80
Thr	Arg	Asn	Gln	Leu 85	Gln	Ala	Ala	Val	Thr 90	Ala	Leu	ràa	Ser	Asn 95	Gly
Ile	Gln	Val	Tyr 100	Gly	Asp	Val	Val	Met 105	Asn	His	Lys	Gly	Gly 110	Ala	Asp
Ala	Thr	Glu 115	Trp	Val	Arg	Ala	Val 120	Glu	Val	Asn	Pro	Ser 125	Asn	Arg	Asn
Gln	Glu 130	Val	Ser	Gly	Asp	Tyr 135	Thr	Ile	Glu	Ala	Trp 140	Thr	Lys	Phe	Asp
Phe 145	Pro	Gly	Arg	Gly	Asn 150	Thr	His	Ser	Asn	Phe 155	Lys	Trp	Arg	Trp	Tyr 160
His	Phe	Asp	Gly	Val 165	Asp	Trp	Asp	Gln	Ser 170	Arg	Gln	Leu	Gln	Asn 175	Arg
Ile	Tyr	ГÀа	Phe 180	Arg	Gly	Asp	Gly	Lys 185	Gly	Trp	Asp	Trp	Glu 190	Val	Asp
Thr	Glu	Asn 195	Gly	Asn	Tyr	Asp	Tyr 200	Leu	Met	Tyr	Ala	Asp 205	Ile	Asp	Met
Asp	His 210	Pro	Glu	Val	Val	Asn 215	Glu	Leu	Arg	Asn	Trp 220	Gly	Val	Trp	Tyr
Thr 225	Asn	Thr	Leu	Gly	Leu 230	Asp	Gly	Phe	Arg	Ile 235	Gly	Ala	Val	Lys	His 240
Ile	Lys	Tyr	Ser	Phe 245	Thr	Arg	Asp	Trp	Leu 250	Thr	His	Val	Arg	Asn 255	Thr
Thr	Gly	Lys	Asn 260	Met	Phe	Ala	Val	Ala 265	Glu	Phe	Trp	Lys	Asn 270	Asp	Ile
Gly	Ala	Ile 275	Glu	Asn	Tyr	Leu	Ser 280	Lys	Thr	Asn	Trp	Asn 285	His	Ser	Val
Phe	Asp 290	Val	Pro	Leu	His	Tyr 295	Asn	Leu	Tyr	Asn	Ala 300	Ser	Arg	Ser	Gly
Gly 305	Asn	Tyr	Asp	Met	Arg 310	Gln	Ile	Phe	Asn	Gly 315	Thr	Val	Val	Gln	Arg 320
His	Pro	Thr	His	Ala 325	Val	Thr	Phe	Val	Asp 330	Asn	His	Asp	Ser	Gln 335	Pro
Glu	Glu	Ala	Leu 340	Glu	Ser	Phe	Val	Glu 345	Glu	Trp	Phe	Lys	Pro 350	Leu	Ala
СЛа	Ala	Leu 355	Thr	Leu	Thr	Arg	360	Gln	Gly	Tyr	Pro	Ser 365	Val	Phe	Tyr
Gly	Asp 370	Tyr	Tyr	Gly	Ile	Pro 375	Thr	His	Gly	Val	Pro 380	Ala	Met	Lys	Ser
385	Ile	Asp	Pro	Ile	Leu 390	Glu	Ala	Arg	Gln	195 195	Tyr	Ala	Tyr	Gly	Lys 400
Gln	Asn	Asp	Tyr	Leu 405	Asp	His	His	Asn	Met 410	Ile	Gly	Trp	Thr	Arg 415	Glu
Gly	Asn	Thr	Ala 420	His	Pro	Asn	Ser	Gly 425	Leu	Ala	Thr	Ile	Met 430	Ser	Asp
Gly	Pro	Gly 435	Gly	Asn	Lys	Trp	Met 440	Tyr	Val	Gly	Arg	Asn 445	Lys	Ala	Gly
Gln	Val 450	Trp	Arg	Asp	Ile	Thr 455	Gly	Asn	Arg	Ser	Gly 460	Thr	Val	Thr	Ile

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Asn Ala Asp Gly Trp Gly Asn Phe Ser Val Asn Gly Gly Ser Val Ser 470 475 Ile Trp Val Asn Asn <210> SEQ ID NO 16 <211> LENGTH: 485 <212> TYPE: PRT <213> ORGANISM: Cytophaga <400> SEQUENCE: 16 Ala Ala Thr Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Val Pro Asn Asp Gly Gln Gln Trp Asn Arg Leu Arg Thr Asp Ala Pro Tyr Leu 20 25 30Ser Ser Val Gly Ile Thr Ala Val Trp Thr Pro Pro Ala Tyr Lys Gly 35 40 45 Thr Ser Gln Ala Asp Val Gly Tyr Gly Pro Tyr Asp Leu Tyr Asp Leu 50  $\phantom{00}$  55  $\phantom{00}$  60 Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys 65 70 75 80 Gly Glu Leu Lys Ser Ala Val Asn Thr Leu His Ser Asn Gly Ile Gln Val Tyr Gly Asp Val Val Met Asn His Lys Ala Gly Ala Asp Tyr Thr 105 Glu Asn Val Thr Ala Val Glu Val Asn Pro Ser Asn Arg Asn Gln Glu 120 Thr Ser Gly Glu Tyr Asn Ile Gln Ala Trp Thr Gly Phe Asn Phe Pro 135 Gly Arg Gly Thr Thr Tyr Ser Asn Phe Lys Trp Gln Trp Phe His Phe 150 Asp Gly Thr Asp Trp Asp Gln Ser Arg Ser Leu Ser Arg Ile Phe Lys Phe Arg Gly Thr Gly Lys Ala Trp Asp Trp Glu Val Ser Ser Glu Asn 185 Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Ile Asp Tyr Asp His Pro Asp Val Val Asn Glu Met Lys Lys Trp Gly Val Trp Tyr Ala Asn Glu Val Gly Leu Asp Gly Tyr Arg Leu Asp Ala Val Lys His Ile Lys Phe Ser Phe Leu Lys Asp Trp Val Asp Asn Ala Arg Ala Ala Thr Gly Lys Glu Met Phe Thr Val Gly Glu Tyr Trp Gln Asn Asp Leu Gly Ala Leu Asn Asn Tyr Leu Ala Lys Val Asn Tyr Asn Gln Ser Leu Phe Asp Ala 280 Pro Leu His Tyr Asn Phe Tyr Ala Ala Ser Thr Gly Gly Gly Tyr Tyr 295 Asp Met Arg Asn Ile Leu Asn Asn Thr Leu Val Ala Ser Asn Pro Thr Lys Ala Val Thr Leu Val Glu Asn His Asp Thr Gln Pro Gly Gln Ser 330 Leu Glu Ser Thr Val Gln Pro Trp Phe Lys Pro Leu Ala Tyr Ala Phe 345

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Ile Leu Thr Arg Ser Gly Gly Tyr Pro Ser Val Phe Tyr Gly Asp Met 360 Tyr Gly Thr Lys Gly Thr Thr Thr Arg Glu Ile Pro Ala Leu Lys Ser 375 Lys Ile Glu Pro Leu Leu Lys Ala Arg Lys Asp Tyr Ala Tyr Gly Thr Gln Arg Asp Tyr Ile Asp Asn Pro Asp Val Ile Gly Trp Thr Arg Glu Gly Asp Ser Thr Lys Ala Lys Ser Gly Leu Ala Thr Val Ile Thr Asp Gly Pro Gly Gly Ser Lys Arg Met Tyr Val Gly Thr Ser Asn Ala Gly Glu Ile Trp Tyr Asp Leu Thr Gly Asn Arg Thr Asp Lys Ile Thr Ile Gly Ser Asp Gly Tyr Ala Thr Phe Pro Val Asn Gly Gly Ser Val Ser Val Trp Val Gln Gln 485 <210> SEO ID NO 17 <211> LENGTH: 486 <212> TYPE: PRT <213> ORGANISM: Bacillus stearothermophilus <400> SEOUENCE: 17 Ala Ala Pro Phe Asn Gly Thr Met Met Gln Tyr Phe Glu Trp Tyr Leu Pro Asp Asp Gly Thr Leu Trp Thr Lys Val Ala Asn Glu Ala Asn Asn Leu Ser Ser Leu Gly Ile Thr Ala Leu Trp Leu Pro Pro Ala Tyr Lys 40 Gly Thr Ser Arg Ser Asp Val Gly Tyr Gly Val Tyr Asp Leu Tyr Asp Leu Gly Glu Phe Asn Gln Lys Gly Thr Val Arg Thr Lys Tyr Gly Thr Lys Ala Gln Tyr Leu Gln Ala Ile Gln Ala Ala His Ala Ala Gly Met Gln Val Tyr Ala Asp Val Val Phe Asp His Lys Gly Gly Ala Asp Gly Thr Glu Trp Val Asp Ala Val Glu Val Asn Pro Ser Asp Arg Asn Gln Glu Ile Ser Gly Thr Tyr Gln Ile Gln Ala Trp Thr Lys Phe Asp Phe Pro Gly Arg Gly Asn Thr Tyr Ser Ser Phe Lys Trp Arg Trp Tyr His Phe Asp Gly Val Asp Trp Asp Glu Ser Arg Lys Leu Ser Arg Ile Tyr 170 Lys Phe Arg Gly Ile Gly Lys Ala Trp Asp Trp Glu Val Asp Thr Glu 185 Asn Gly Asn Tyr Asp Tyr Leu Met Tyr Ala Asp Leu Asp Met Asp His Pro Glu Val Val Thr Glu Leu Lys Asn Trp Gly Lys Trp Tyr Val Asn Thr Thr Asn Ile Asp Gly Phe Arg Leu Asp Ala Val Lys His Ile Lys

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225					230					235					240
Phe	Ser	Phe	Phe	Pro 245	Asp	Trp	Leu	Ser	Tyr 250	Val	Arg	Ser	Gln	Thr 255	Gly
ГÀв	Pro	Leu	Phe 260	Thr	Val	Gly	Glu	Tyr 265	Trp	Ser	Tyr	Asp	Ile 270	Asn	Lys
Leu	His	Asn 275	Tyr	Ile	Thr	Lys	Thr 280	Asn	Gly	Thr	Met	Ser 285	Leu	Phe	Asp
Ala	Pro 290	Leu	His	Asn	Lys	Phe 295	Tyr	Thr	Ala	Ser	300 TÀs	Ser	Gly	Gly	Ala
Phe 305	Asp	Met	Arg	Thr	Leu 310	Met	Thr	Asn	Thr	Leu 315	Met	ГÀа	Asp	Gln	Pro 320
Thr	Leu	Ala	Val	Thr 325	Phe	Val	Asp	Asn	His 330	Asp	Thr	Glu	Pro	Gly 335	Gln
Ala	Leu	Gln	Ser 340	Trp	Val	Asp	Pro	Trp 345	Phe	Lys	Pro	Leu	Ala 350	Tyr	Ala
Phe	Ile	Leu 355	Thr	Arg	Gln	Glu	Gly 360	Tyr	Pro	Cys	Val	Phe 365	Tyr	Gly	Asp
Tyr	Tyr 370	Gly	Ile	Pro	Gln	Tyr 375	Asn	Ile	Pro	Ser	Leu 380	Lys	Ser	Lys	Ile
Asp 385	Pro	Leu	Leu	Ile	Ala 390	Arg	Arg	Asp	Tyr	Ala 395	Tyr	Gly	Thr	Gln	His 400
Asp	Tyr	Leu	Asp	His 405	Ser	Asp	Ile	Ile	Gly 410	Trp	Thr	Arg	Glu	Gly 415	Val
Thr	Glu	Lys	Pro 420	Gly	Ser	Gly	Leu	Ala 425	Ala	Leu	Ile	Thr	Asp 430	Gly	Pro
Gly	Gly	Ser 435	Lys	Trp	Met	Tyr	Val 440	Gly	Lys	Gln	His	Ala 445	Gly	Lys	Val
Phe	Tyr 450	Asp	Leu	Thr	Gly	Asn 455	Arg	Ser	Asp	Thr	Val 460	Thr	Ile	Asn	Ser
Asp 465	Gly	Trp	Gly	Glu	Phe 470	Lys	Val	Asn	Gly	Gly 475	Ser	Val	Ser	Val	Trp 480
Val	Pro	Arg	Lys	Thr 485	Thr										

The invention claimed is:

- 1. An isolated variant alpha-amylase, comprising two or more alterations at positions corresponding to positions 163, 188, 205, 208, and 209 of the polypeptide of SEQ ID NO: 2,
  - (a) the variant has a sequence identity to SEQ ID NO: 8 of 50at least 90% and less than 100%;
  - (b) each alteration is independently a substitution, deletion or insertion; and
  - (c) the variant has alpha-amylase activity.
- identity to sequence identity to SEQ ID NO: 8.
- 3. The variant of claim 1, which has at least 99% sequence identity to sequence identity to SEQ ID NO: 8.
- 4. The variant of claim 1, wherein the alterations at posi-  $_{60}$ tions 163, 188, 205, 208, and 209 are substitutions.
- 5. The variant of claim 1, wherein the alterations are selected among: X163Q,N, X188N, X205N, X208Y and X209N,S.
- 6. The variant of claim 1, further comprising one or more 65 alterations selected from the group consisting of a deletion at a position corresponding to positions 183 and 184 and a

- substitution at a position corresponding to the positions selected from the group consisting of 186, 193, 195, 202, 206, 214, 244, 452, 474 and 475.
- 7. The variant of claim 1, which further comprises one or more alterations selected from the group of X181\*+X182\*, X182\*+X183\*, X183\*+X184\*, X185K, X167W, X202L/I/ T, X203Y, X167W+X168E+X169E+X170R, X51T+ X109G+X203Y, X109G+X203Y, X189W, X189W+x190E+ x193T, X190E, X193T, X303K, X303K+x305R+x306D+ 2. The variant of claim 1, which has at least 95% sequence 55 X409N+X432N+X434D, X305R, X306D, X409N, X432N, and X434D.
  - 8. The variant of claim 1, which further comprises one or more alterations selected from the group consisting of A113E, N116V, V117F, L118K, A119V, V120I, N123D, N126D, N128T, Q129K, G133E, D134P, Y135F, T136E, A139G, D144T, N150D, T151Q, D154S, R158N, W159S, Y160E, V165T, W167F, Q169A, S170K, R171G, Q172\*, F173E, Q174R, N175T, R176G, I177V, Y178F, K179R, F180I, R181A, D183E, G184N, A186K, W189E, E190N, S193T/D, N195F, Y203F, V206I, E212D, V214R, and N215R.

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9. The variant of claim 1 selected from the group consisting of:

D188N+D209S;

D163N+D188N+D209S;

D163N+D188N+D205N+D209S;

D163N+D188N+D205N+M208F+D209S;

D207N+D209S;

D163N+D207N+D209S;

D163N+D188N+D207N+D209S;

D163N+D188N+D199N+D207N+D209S;

D163N+D188N+D199N+D205N+D207N+D209S;

D163N+D188N+D199N+D205N+M208F+D207N+ D209S:

D163N+R181A+G182N+G186N+D188N+D205N+ D209S;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+D209S;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S;

D163N+R181A+G182N+D188N+D199N+D205N+ M208F+D207N+D209S;

D163N+R181A+G182N+K185T+D188N+D199N+ D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+G186N+D188N+ D199N+D205N+M208F+D207N+D209S;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+N409D+ D432N+A434P;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+K242P+S244W+H408W+ N409D+D432N+A434P;

N128W+D163N+R181A+G182N+K185T+G186N+ D188N+D205N+M208F+D209S+K242P+S244W;

D163N+R181A+G182N+K185T+G186N+D188N+ D205N+M208F+D209S+V238I+K242P+S24 4W; and D163Q+D188N+M208F+D209S+K242P+S244W.

10. The variant of claim 1 selected from the group consisting of:

D188N+D209S,

D163N+D188N+D209S,

D163N+D188N+D205N+D209S,

D163N+D188N+D205N+M208F+D209S,

D207N+D209S,

D163N+D207N+D209S.

D163N+D188N+D207N+D209S.

D163N+D188N+D199N+D207N+D209S,

D163N+D188N+D199N+D205N+D207N+D209S,

D163N+D188N+D199N+D205N+M208F+D207N+ D209S, 88

D163N+R181A+G182N+G186N+D188N+D205N+ D209S.

D163N+R181A+G182N+K185T+A186N+D188N+D205N+D209S.

D163N+R181A+G182N+K185T+A186N+D188N+ D205N+M208F+D209S.

D163N+R181A+G182N+D188N+D199N+D205N+ M208F+D207N+D209S,

D163N+R181A+G182N+K185T+D188N+D199N+ D205N+M208F+D207N+D209S, and

D163N+R181A+G182N+K185T+A186N+D188N+ D199N+D205N+M208F+D207N+D209S.

11. The variant of claim 1 selected from the group consisting of:

D207N+D186N,

D207N+D186N+D162N,

D207N+D186N+D162N+D203N,

D207N+D186N+D162N+D203N+M206Y,

D207N+D186N+D162N+D203N+M206Y+D105N,

D207N+A184K+T187E,

D207N+A184K+T187E+D186N,

D207N+A184K+T187E+D186N+D162N,

D207N+A184K+T187E+D186N+D162N+D203N,

D207N+A184K+T187E+D186N+D162N+D203N+ M206Y, and

D207N+Á184K+T187E+D186N+D162N+D203N+ M206Y+D105N.

12. A detergent composition comprising a variant of claim 1 and a surfactant.

13. A method of producing liquefied starch, comprising liquefying a starch-containing material with a variant of claim

14. A process of producing a fermentation product, comprising

 a. liquefying a starch-containing material with a variant of claim 1 to produce a liquefied mash;

b. saccharifying the liquefied mash to produce fermentable sugars; and

c. fermenting the fermentable sugars in the presence of a fermenting organism.

15. A process of producing a fermentation product, comprising contacting a starch substrate with a variant of claim 1, a glucoamylase, and a fermenting organism.

16. An isolated variant alpha-amylase comprising at least two substitutions at positions corresponding to positions 186 and 207 of the mature polypeptide of SEQ ID NO: 8, wherein

(a) the variant has a sequence identity to SEQ ID NO: 8 of at least 95% and less than 100%; and

(b) the variant has alpha-amylase activity.

\* \* \* \* \*

# UNITED STATES PATENT AND TRADEMARK OFFICE CERTIFICATE OF CORRECTION

PATENT NO. : 9,273,297 B2

APPLICATION NO. : 14/525877
DATED : March 1, 2016
INVENTOR(S) : Andersen

It is certified that error appears in the above-identified patent and that said Letters Patent is hereby corrected as shown below:

In the claims

Column 88, line 44-50, Claim 16 should read

--An isolated variant alpha-amylase comprising at least two substitutions at positions corresponding to positions 186 and 207 of the polypeptide of SEQ ID NO: 8, wherein

- (a) the variant has a sequence identity to SEQ ID NO: 8 of at least 95% and less than 100%; and
  - (b) the variant has alpha-amylase activity.--

Signed and Sealed this Thirteenth Day of September, 2016

Michelle K. Lee

Michelle K. Lee

Director of the United States Patent and Trademark Office